



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification: C12N 15/10, C12Q 1/68	A2	(11) International Publication Number: WO 00/15779 (43) International Publication Date: 23 March 2000 (23.03.2000)
(21) International Application Number: PCT/US99/21291 (22) International Filing Date: 15 September 1999 (15.09.1999) (30) Priority Data: 60/100,327 15 September 1998 (15.09.1998) US (60) Parent Application or Grant YALE UNIVERSITY [/]; O. LIZARDI, Paul, M. [/]; O. PABST, Patrea, L.; O.	Published	
(54) Title: MOLECULAR CLONING USING ROLLING CIRCLE AMPLIFICATION (54) Titre: CLONAGE MOLECULAIRE A AMPLIFICATION SELON LE MODELE DU CERCLE ROULANT		
(57) Abstract <p>Disclosed are reagents and a method for efficient in vitro molecular cloning of nucleic acid molecules of interest. Because the method is entirely in vitro, it can be automated and scaled-up in ways that are not possible in cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process. The amplification is rapid and efficient since it involves a single, isothermal reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be used for any purpose and in any manner that nucleic acid cloned or amplified by known methods can be used. This includes sequencing, probing, restriction analysis, subcloning, transcription, hybridization or denaturation analysis, further amplified, and storage for future use or analysis.</p> (57) Abrégé <p>L'invention concerne des réactifs et un procédé permettant d'assurer un clonage moléculaire in vitro de molécules d'acides nucléiques déterminées. Etant entièrement in vitro, le procédé peut être automatisé et étendu selon des modalités qu'il est impossible de mettre en oeuvre avec le clonage moléculaire à base cellulaire. Le procédé consiste à insérer la molécule d'acide nucléique concernée dans un vecteur linéaire, de manière à former un vecteur circulaire dans lequel un brin est continu et l'autre brin est discontinu. Le brin continu est ensuite amplifié selon le modèle du cercle roulant, ce qui permet d'amplifier par la même occasion la molécule d'acide nucléique. L'amplification est rapide et efficace car elle fait intervenir une réaction isothermique unique qui assure la réplication exponentielle des séquences du vecteur. Le procédé peut être automatisé lorsque plusieurs réactions sont conduites simultanément dans une zone de taille réduite. L'acide nucléique amplifié peut être utilisé à toute fin et de façon quelconque dans l'éventail d'utilisation connu de l'acide nucléique cloné ou amplifié, y compris le séquençage, la détermination de sondes, l'analyse de restriction, le sous-clonage, la transcription, l'analyse d'hybridation ou de dénaturation, le complément d'amplification et le stockage pour utilisation ou analyse ultérieure.</p>		

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<p>(21) International Application Number: PCT/US99/21291 (22) International Filing Date: 15 September 1999 (15.09.99) (30) Priority Data: 60/100,327 15 September 1998 (15.09.98) US (71) Applicant: YALE UNIVERSITY [US/US]; 451 College Street, New Haven, CT 06511 (US). (72) Inventor: LIZARDI, Paul, M.; 350 Mountain Road, Hamden, CT 06514 (US). (74) Agents: PABST, Patrea, L. et al.; Arnall Golden & Gregory, LLP, 2800 One Atlantic Center, 1201 West Peachtree Street, Atlanta, GA 30309-3450 (US).</p>		<p>(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published Without international search report and to be republished upon receipt of that report.</p>
<p>(54) Title: MOLECULAR CLONING USING ROLLING CIRCLE AMPLIFICATION</p>		
<p>(57) Abstract</p> <p>Disclosed are reagents and a method for efficient <i>in vitro</i> molecular cloning of nucleic acid molecules of interest. Because the method is entirely <i>in vitro</i>, it can be automated and scaled-up in ways that are not possible in cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process. The amplification is rapid and efficient since it involves a single, isothermic reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be used for any purpose and in any manner that nucleic acid cloned or amplified by known methods can be used. This includes sequencing, probing, restriction analysis, subcloning, transcription, hybridization or denaturation analysis, further amplified, and storage for future use or analysis.</p>		

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**MOLECULAR CLONING USING
ROLLING CIRCLE AMPLIFICATION
BACKGROUND OF THE INVENTION**

The disclosed invention is generally in the field of molecular cloning and nucleic acid amplification, and specifically involves rolling circle replication of nucleic acid molecules inserted into circular vectors.

DNA molecular cloning is routinely carried out using plasmid, phage, or viral vectors that replicate inside cells. A method, in which individual DNA molecules are cloned in solution by serial dilution and subsequent PCR amplification from tubes containing single molecules has been described (Lukyanov *et al.*, *Nucleic Acid Research* 24:2194-2195 (1996)). A method has also been described for cloning RNA populations derived from single RNA molecules in an immobilized medium (Chetverina and Chetverin, *Nucleic Acids Research* 21:2349-2353 (1993)). While both of these methods allow *in vitro* cloning, neither is practical for high throughput cloning.

Velculescu *et al.*, *Science* 270:484-487 (1995), have described a method for the quantitative cataloguing and comparison of expressed genes in normal, developmental, and disease states. The method, termed serial analysis of gene expression (SAGE), is based in the use of relatively short sequence tags for the unique identification of cDNAs derived from mRNA transcripts. While this method is very powerful, the study of low-abundance mRNAs can require several months of work in order to obtain sufficient sequence information for a complete SAGE analysis of one tissue sample. Thus, there is a need for a method to obtain the sequence of sequence tags more rapidly.

It is therefore an object of the present invention to provide a more efficient method of *in vitro* molecular cloning.

It is also an object of the present invention to provide vectors and kits useful for *in vitro* cloning.

It is also an object of the present invention to provide an automated method molecular cloning.

It is also an object of the present invention to provide a more efficient method of sequential analysis of gene expression.

BRIEF SUMMARY OF THE INVENTION

Disclosed are reagents and a method for efficient *in vitro* molecular cloning of nucleic acid molecules of interest. Because the method is entirely *in vitro*, it can be automated and scaled-up in ways that are not possible in cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process. The amplification is rapid and efficient since it involves a single, isothermal reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be used for any purpose and in any manner that nucleic acid cloned or amplified by known methods can be used. This includes sequencing, probing, restriction analysis, subcloning, transcription, hybridization or denaturation analysis, further amplified, and storage for future use or analysis.

The insertion reaction involves insertion of a double-stranded nucleic acid molecule into a double-stranded linear vector to produce a double-stranded circular vector. The use of circular vectors facilitates the selection of molecules that have successfully incorporated inserts. The amplification reaction involves rolling circle replication of a single-stranded circular nucleic acid molecule.

A key feature of the method, which facilitates double-stranded insertion followed by single-stranded amplification, is formation of the circular vector in such a way that one of its strands is a closed circular strand (that is, continuous) while the other strand is not a closed circular strand (that is, it has a nick, a gap, an overlap, or is otherwise discontinuous). This feature is most useful, and most effectively accomplished, when, by operation of the method, the closed strand and the open strand are predetermined; that is, when a particular strand of the vector is selectively left discontinuous.

5 With rolling circle replication, amplification takes place not in cycles,
but in a continuous, isothermal replication. This makes amplification less
complicated and much more consistent in output. A single round of rolling
10 circle replication results in a large amplification of the circular vector, orders
5 of magnitude greater than a single cycle of PCR replication and other
amplification techniques in which each cycle is limited to a doubling of the
number of copies of a target sequence.

15 Following amplification, the amplified nucleic acid can be used for
any purpose. Numerous methods for the use and manipulation of cloned or
10 isolated nucleic acid are known and can be applied to nucleic acid amplified
in the present method. For example, the nucleic acid can be sequenced,
20 probed, subjected to restriction analysis, subcloned, transcribed, subjected to
hybridization or denaturation analysis, further amplified, or stored.
Diagnostic methods, such as sequencing and probing for specific sequences,
25 are preferred.

Libraries of cloned nucleic acids formed by the disclosed method can
be screened using any of the methods used for screening conventional
libraries. For example, cDNA libraries made using the disclosed method can
30 be analyzed using conventional screens. Libraries can also be used for *in*
20 *situ* transcription to generate RNA colonies, which can then be analyzed (*in*
situ or in replicas) by appropriate screens, such as aptamer screens or
35 ribozyme activity screens. Libraries can also be screened by *in situ*
translation on array replicas (see, for example, Saris *et al.*, *Nucleic Acids Res.*
10:4831-4843 (1982)). Libraries can also be screened by *in situ* coupled
25 transcription-translation systems, and subsequent catalytic activity assays for
the analysis of mutagenized enzymes. Libraries can be screened and
40 cataloged by sequencing and use of the data for the analysis of cDNA
abundancies, which is useful for RNA profiling and serial analysis of gene
expression (SAGE; Velculescu *et al.*, *Science* 270:484-487 (1995)).

30 BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a diagram of examples of various forms of linear vectors
and the circular vectors, having one continuous strand and one discontinuous
50 strand, that result upon insertion of a nucleic acid molecule.

5 Figure 2 is a diagram of an example of the disclosed method where the linear vector includes an non-ligatable nick in one of its strands. Upon ligation to form a circular vector, the discontinuous strand is separated from the continuous strand by binding the biotin moiety at the nick to an
10 5 immobilized streptavidin moiety.

Figure 3 is a diagram of an example of the disclosed method where the linear vector has an overlap. Upon ligation to form a circular vector (with a Y tail), the discontinuous strand is separated from the continuous strand by ligating one end of the discontinuous strand to an immobilized
15 10 nucleic acid probe. This ligation is mediated by hybridization between the single-stranded extension of the discontinuous strand and the probe.

Figure 4 is a diagram of an example of the disclosed method where the linear vector is a linker having sticky ends compatible with sticky ends formed by restriction digestion of PCR primer sequences incorporated at the
20 25 15 ends of PCR amplified DNA. The linker facilitates circularization of the PCR amplified DNA. The linker has one non-ligatable end which, upon ligation, results in a circular molecule with one continuous strand and one discontinuous strand. The discontinuous strand is separated from the continuous strand by binding the biotin moiety at the nick to an immobilized
30 20 streptavidin moiety.

DETAILED DESCRIPTION OF THE INVENTION

35 Disclosed are reagents and a method for efficient *in vitro* molecular cloning of nucleic acid molecules of interest. Because the method is entirely *in vitro*, it can be automated and scaled-up in ways that are not possible in
40 25 cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process.
45 30 The amplification is rapid and efficient since it involves a single, isothermal reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be

5 used for any purpose and in any manner that nucleic acid cloned or amplified
by known methods can be used. This includes sequencing, probing,
restriction analysis, subcloning, transcription, hybridization or denaturation
analysis, further amplified, and storage for future use or analysis.

10 5 The insertion reaction involves insertion of a double-stranded nucleic
acid molecule into a double-stranded linear vector to produce a double-
stranded circular vector. The use of circular vectors facilitates the selection
15 of molecules that have successfully incorporated inserts. The amplification
reaction involves rolling circle replication of a single-stranded circular
20 10 nucleic acid molecule. In its most useful forms, the disclosed method
involves insertion of nucleic acid molecules of interest into a vector and
separate amplification of the resulting recombinant vectors to produce
separate nucleic acid "colonies," each representing a clonal population of
nucleic acid sequences present in founding vector of that "colony."

25 15 A key feature of the method, which facilitates double-stranded
insertion followed by single-stranded amplification, is formation of the
circular vector in such a way that one of its strands is a closed circular strand
30 (that is, continuous) while the other strand is not a closed circular strand (that
is, it has a nick, a gap, an overlap, or is otherwise discontinuous). This
20 feature is most useful, and most effectively accomplished, when, by
operation of the method, the closed strand and the open strand are
predetermined; that is, when a particular strand of the vector is selectively
35 left discontinuous.

With rolling circle replication, amplification takes place not in cycles,
25 but in a continuous, isothermal replication. This makes amplification less
40 complicated and much more consistent in output. A single round of rolling
circle replication results in a large amplification of the circular vector, orders
of magnitude greater than a single cycle of PCR replication and other
45 amplification techniques in which each cycle is limited to a doubling of the
30 number of copies of a target sequence.

The present method is an alternative to traditional molecular cloning
involving clonal amplification in cells harnessing the natural nucleic acid
50 replication in, and growth and division of, the cells. The present method has

WO 00/15779

5 several advantages over this traditional method. First, it is much more rapid.
Traditional molecular cloning usually requires at least twelve hours of cell
growth in the best cell-based cloning methods to produce a million copies of
a single vector. In contrast, the present method allows production of millions
10 or billions of copies of a single vector in only 60 to 90 minutes.

5 Once a clonal culture or colony of cells is grown, there is still the
problem of separating the amplified nucleic acid from the cell and all the
cellular components. Although numerous methods have been devised over
15 the years for such purification, they remain both time consuming and
ineffective (that is, cellular contaminants remain with the isolated nucleic
20 acid). Generally, the amount of time for the purification and the level of
purity obtain from nucleic acid isolation methods is proportional: more time,
more purity; less time, less purity. In contrast, the present method
accomplishes clonal amplification in an uncomplicated mix of just a few
25 well-defined components: the vector, one or two types of primers,
nucleotides, and polymerase. The purification of the amplified nucleic acid
is correspondingly simplified. Most significantly, the amplification reaction
in present method need not result in a complex mixture of nucleic acids as is
30 true of cell-based molecular cloning.

20 The present method also has advantages over cyclic amplification
methods such as the polymerase chain reaction (PCR). Rolling circle
replication is more rapid and has higher yields than PCR. Significantly, the
35 products of rolling circle replication are tandemly repeated amplicons of
double-stranded DNA. These amplicons, if carried as contaminants to any
25 surface or vessel, are unable to seed new rolling circle replication reactions.
In other words, the amplified DNA is non-contaminating because it is a
40 replication dead-end. By contrast, PCR or SDA amplicons are potentially
contaminating.

45 Following amplification, the amplified nucleic acid can be used for
any purpose. Numerous methods for the use and manipulation of cloned or
30 isolated nucleic acid are known and can be applied to nucleic acid amplified
in the present method. For example, the nucleic acid can be sequenced,
50 probed, subjected to restriction analysis, subcloned, transcribed, subjected to

5 hybridization or denaturation analysis, further amplified, or stored.
Diagnostic methods, such as sequencing and probing for specific sequences,
are preferred.

10 The nucleotide sequence of the amplified sequences can be
5 determined either by conventional means or by primer extension sequencing
of amplified target sequence. One preferred form of sequencing for use with
amplified sequences produced with the disclosed method is nanosequencing
or single-nucleotide extension sequencing. Nanosequencing methods are
15 described below and by Jalanko *et al.*, *Clinical Chemistry* 38:39-43 (1992);
Nikiforov *et al.*, *Nucleic Acids Research* 22:4167-4175 (1994); and
10 Kobayashi *et al.*, *Molecular and Cellular Probes* 9:175-182 (1995).

20 Two forms of sequencing that can be used with the disclosed method
are described in PCT Application WO 97/20948. One is single nucleotide
primer extension sequencing involving interrogation of a single nucleotide in
25 an amplified target sequence by incorporation of a specific and identifiable
nucleotide based on the identity of the interrogated nucleotide. The other is
degenerate probe primer extension sequencing involving sequential addition
of degenerate probes to an interrogation primer hybridized to amplified
30 target sequences.

20 Libraries of cloned nucleic acids formed by the disclosed method can
be screened using any of the methods used for screening conventional
libraries. For example, cDNA libraries made using the disclosed method can
35 be analyzed using conventional screens. Libraries can also be used for *in*
situ transcription to generate RNA colonies, which can then be analyzed (*in*
25 *situ* or in replicas) by appropriate screens, such as aptamer screens or
ribozymic activity screens. Libraries can also be screened by *in situ*
40 translation on array replicas (see, for example, Saris *et al.*, *Nucleic Acids Res.*
10:4831-4843 (1982)). Libraries can also be screened by *in situ* coupled
transcription-translation systems, and subsequent catalytic activity assays for
45 the analysis of mutagenized enzymes. Libraries can be screened and
30 cataloged by sequencing and use of the data for the analysis of cDNA
abundancies, which is useful for RNA profiling and serial analysis of gene
expression (SAGE; Velculescu *et al.*, *Science* 270:484-487 (1995)).
50

5 One embodiment of the disclosed method is a method of isolating
and amplifying a nucleic acid molecule, where the method involves:

10 (a) ligating a nucleic acid molecule into a linear vector to form a
circular vector including the vector and the nucleic acid molecule, where the
5 linear vector is a double-stranded linear nucleic acid including two nucleic
acid strands, where the second strand of the circular vector is discontinuous,
and where the first strand in the circular vector is a closed circular strand,
15 and

(b) amplifying the first strand by rolling circle replication to form
10 tandem sequence DNA, where the amplification results in amplification of
the nucleic acid molecule in the first strand.

20 The method can be practiced and expanded in several ways. For
example, the second strand of the linear vector can contain at least one nick,
where the nick cannot be ligated. The linear vector can be designed such that
15 either the 5' or the 3' end of the second strand of the linear vector cannot be
ligated. The linear vector can be designed such that the second strand of the
linear vector contains at least one gap or overlap. The method can be
extended to include, following ligation and prior to amplification, separation
30 of the first strand from the second strand. The second strand of the vector
can include an affinity tag. In this case, the first strand can be separated from
the second strand by binding the affinity tag to a substrate, denaturing the
first and second strands prior to, simultaneous with, or following binding,
35 and separating the first strand from the substrate.

The second strand of the linear vector can also be designed to contain
25 at least one overlap, where part of the overlapping portions of the second
strand are complementary, and where the 3' end of the overlap extends
beyond the part of the overlapping portions that are complementary. In this
case, the first strand can be separated from the second strand by ligating one
40 end of the second strand to a nucleic acid molecule coupled to a substrate,
denaturing the first and second strands following ligation of the second
30 strand, and separating the first strand from the substrate.

The method can also be practiced such that step (a) involves ligating
50 a plurality of nucleic acid molecules into a plurality of linear vectors in a

5 single reaction to form a plurality of circular vectors, each circular vector
containing at least one nick, gap, or overlap in the second strand, such that
step (b) involves amplifying the first strand of the plurality of circular
10 5 vectors, and such that, prior to amplification, the ligation reaction is divided
to produce a plurality of separate amplification reactions. The method can be
extended to include making a replica of the amplification reactions. The
15 replica of the amplification reactions can be made by contacting the
amplification reactions with a surface to which nucleic acids can bind. The
replica of the amplification reactions can also be made by transferring part of
10 each amplification reaction to form a replica amplification reaction.

20 In the method, the ligation reaction can be divided by spreading the
ligation reaction onto a surface to form a spread, where the separate
amplification reactions are the locations of circular vectors on the surface
after spreading. In this case, a replica of the amplification reactions can be
25 15 made by contacting the spread with a second surface to which nucleic acids
can bind.

30 Any number or all of the amplification reactions can be ordered as an
array of reaction droplets or in an array of reaction vessels. In this case,
following amplification, all or part of the contents of any number or all the
20 individual reaction droplets or reaction vessels are transferred by one to one
mapping to a new set of reaction droplets or reaction vessels.

35 The method can be further extended by, following amplification,
determining the presence of amplified nucleic acid in the amplification
reactions, and transferring all or a part of the contents of the amplification
25 reactions containing amplified nucleic acid reaction to a new set of reaction
droplets or reaction vessels.

40 Replicas of the amplification reactions can be made by contacting the
amplification reactions with a surface treated with an affinity target capable
of binding an affinity tag, where the amplified nucleic include affinity tags
45 30 incorporated during amplification, such that a portion of each amplification
reaction is transferred to the surface. The affinity tag is preferably biotin and
the affinity target is preferably streptavidin. The affinity tag is also
50 preferably a reactive moiety and the affinity target is preferably a

5 corresponding reactive moiety, such that a chemical reaction between the
affinity tag and the affinity target results in the amplified nucleic acid being
covalently coupled to the surface. In this case, it is preferred that the affinity
target is phenylene diisothiocyanate, disuccinimidylcarbonate,
10 5 disuccinimidylloxolate or dimethylsuberimide and the affinity tag is a
reactive amine.

A replica of the amplification reactions can also be made by
transferring part of each amplification reaction to form a replica
15 amplification reaction. Following amplification, all or part of the contents of
any number or all of the reaction droplets or reaction vessels can be
transferred and combined to create one or more sets of pooled reactions. The
20 amplification reactions can also be arranged on the surface of a substrate.
Preferred substrates include acrylamide, cellulose, nitrocellulose,
polystyrene, polyethylene vinyl acetate, polypropylene, polymethacrylate,
25 15 polyethylene, polyethylene oxide, glass, polysilicates, polycarbonates, teflon,
fluorocarbons, nylon, silicon rubber, polyanhydrides, polyglycolic acid,
polylactic acid, polyorthoesters, polypropylfumerate, collagen,
glycosaminoglycans, polyamino acids, chemical resistant metals, or
30 corrosion resistant metals.

20 The ligation reactions can also be diluted prior to division of the
ligation reaction into amplification reactions such that, on average, each
amplification reaction contains a single circular vector. A sample of each
35 amplification reaction can also be collected. Nucleic acid molecules in the
amplification reactions or in the collected samples can also be detected or
25 sequenced.

40 In the disclosed method, rolling circle replication can be primed by
the second strand on the circular vector or by a rolling circle replication
primer. The tandem sequence DNA formed after amplification can itself be
amplified by strand displacement replication to form secondary tandem
45 30 sequence DNA. This secondary tandem sequence DNA can also be
amplified by strand displacement replication to form tertiary tandem
sequence DNA. Strand displacement replication of the tandem sequence can
be primed by a strand displacement primer.
50

5 The method can also include detection of one or more amplified
nucleic acid molecules in one or more of the amplification reactions. In a
preferred embodiment, the nucleic acid molecules can be derived from
cDNA generated by suppression subtractive hybridization. In another
10 5 preferred embodiment, the plurality of nucleic acid molecules can be all
derived from the same source.

 Detection can be accomplished by, following amplification, creating
a replica of the amplification reactions, contacting the amplification reactions
15 with a first set of labeled nucleic acid probes and the replica amplification
reactions with a second set of labeled nucleic acid probes, and comparing the
20 pattern of hybridization of the first set of probes to the pattern of
hybridization of the second set of probes, where differences in the patterns of
hybridization indicate differences in the probe sets. Following detection
amplification reactions that hybridize to the first set of probes but not to the
25 second set of probes, amplification reactions that hybridize to the second set
of probes but not to the first set of probes, amplification reactions that
hybridize to the both sets of probes, or amplification reactions that do not
hybridize to either set of probes can be selected for isolation or further
30 analysis.

20 Another embodiment of the disclosed method is an *in vitro* method of
cloning nucleic acid molecules, where the method involves

- 35 (a) dividing a nucleic acid sample to produce a plurality of separate
amplification reactions,
 (b) amplifying nucleic acid molecules in the amplification reactions,
25 (c) making a replica of the amplification reactions,
 (d) testing nucleic acid molecules in either the amplification
40 reactions or the replica amplification reactions to identify nucleic acid
molecules of interest, and
 (e) retrieving the identified nucleic acid molecules of interest from
45 30 the corresponding amplification reactions or replica amplification reactions
that were not tested.

 In this embodiment, the nucleic acid sample can be divided by
50 spreading the sample onto a surface to form a spread, such that the separate

5 amplification reactions are the locations of circular vectors on the surface after spreading. The replica of the amplification reactions can be made by contacting the spread with a second surface to which nucleic acids can bind.

Another embodiment of the disclosed method is a method of isolating

10 5 and amplifying nucleic acid molecules, where the method involves

(a) ligating a plurality of nucleic acid molecules into a plurality of linear vectors in a single reaction to form a plurality of circular vectors, each circular vector including a vector and a nucleic acid molecule, where the linear vectors are double-stranded linear nucleic acid comprising two nucleic acid strands, where the circular vectors each contain at least one nick, gap, or overlap in the second strand, and where the first strand in each circular vector is a closed circular strand,

(b) separating the first strands from the second strands,

(c) diluting and dividing the first strands to produce a plurality of separate amplification reactions that, on average, each contain a single circular vector,

(d) amplifying the first strands of the plurality of circular vectors by rolling circle replication to form tandem sequence DNA, where the amplification results in amplification of the nucleic acid molecules in the first strands.

The tandem sequence DNA formed after amplification can itself be amplified by strand displacement replication to form secondary tandem sequence DNA. This secondary tandem sequence DNA can also be amplified by strand displacement replication to form tertiary tandem sequence DNA.

Another embodiment of the disclosed method is a method of isolating and amplifying a nucleic acid molecule, where the method involves

(a) ligating a nucleic acid molecule into a linear vector to form a circular vector comprising the vector and the nucleic acid molecule, where the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, where the circular vector contains at least one nick in the second strand, and where the first strand in the circular vector is a closed circular strand,

(b) amplifying the first strand, where the amplification results in amplification of the nucleic acid molecule in the first strand.

Also disclosed is a kit for isolating and amplifying nucleic acid molecules, where the kit includes

(a) a linear vector where the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, and where

(1) the linear vector contains at least one nick, where the nick cannot be ligated,

(2) either the 5' or the 3' end of the second strand of the linear vector cannot be ligated,

(3) the second strand of the linear vector contains at least one gap,

(4) the second strand of the linear vector contains at least one overlap, or

(5) any combination of (1), (2), (3) or (4);

(b) a rolling circle replication primer, where the rolling circle replication primer is complementary to a portion of the first strand of the linear vector; and

(c) a strand displacement primer, where the strand displacement primer matches a portion of the first strand of the linear vector.

Also disclosed is a linear vector where the linear vector is a double-stranded linear nucleic acid made up of two nucleic acid strands, where the second strand of the linear vector contains an affinity tag, and where

(1) the linear vector contains at least one nick, where the nick cannot be ligated,

(2) either the 5' or the 3' end of the second strand of the linear vector cannot be ligated,

(3) the second strand of the linear vector contains at least one gap,

(4) the second strand of the linear vector contains at least one overlap, or

(5) any combination of (1), (2), (3) or (4).

Also disclosed is a linear vector where the linear vector is a double-stranded linear nucleic acid made up of two nucleic acid strands, where the second strand of the linear vector contains at least one overlap, part of the overlapping portions of the second strand are complementary, and the 3' end of the overlap extends beyond the part of the overlapping portions that are complementary.

I. Materials

The disclosed method makes use of linear vectors in which nucleic acid molecules of interest can be inserted to form circular vectors. The circular vectors contain one continuous strand and one discontinuous strand. The discontinuous strand may include an affinity tag which, by interaction with an affinity substrate, can facilitate separation of the continuous strand from the discontinuous strand. The continuous strand of the circular vector is amplified by rolling circle replication to form tandem sequence DNA (TS-DNA). Rolling circle replication is primed by a rolling circle replication primer complementary to a sequence in the continuous strand. The tandem sequence DNA itself may be amplified by strand displacement replication, to form secondary tandem sequence DNA, using strand displacement primers (complementary to a sequence in the tandem sequence DNA). The secondary tandem sequence DNA may also be amplified by strand displacement replication, to form tertiary tandem sequence DNA, using strand displacement primers or the rolling circle replication primers (complementary to a sequence in the secondary tandem sequence DNA). Multiple amplification reactions can be carried out in parallel, preferably in arrays or as spreads of diluted vectors on surfaces or embedded in agarose. The resulting "colonies" of amplified DNA represent molecular clones of the progenitor circular vectors with an inserted nucleic acid molecule. Collectively, such colonies form a library of cloned nucleic acid molecules that can be replica plated or arrayed, stored, and screened. These materials are described in detail below.

A. Nucleic Acid Molecules

The disclosed method can be used to clone or amplify any nucleic acid molecule of interest. The nucleic acid molecules can come from any

5 source such as a cellular or tissue nucleic acid sample, a subclone of a
previously cloned fragment, mRNA, chemically synthesized nucleic acid,
genomic nucleic acid samples, nucleic acid molecules obtained from nucleic
acid libraries, specific nucleic acid molecules, and mixtures of nucleic acid
10 5 molecules. The disclosed method is particularly suited to producing libraries
of cloned nucleic acid molecules starting with a complex mixture of nucleic
acid molecules to be represented in the library. For example, cDNA can be
produced from all of the mRNA in a cellular sample and used to make a
15 cDNA library, or a library of genomic DNA can be produced from a
20 genomic nucleic acid sample.

In the method, the nucleic acid molecule is inserted into a double-
20 stranded linear vector. Preferably the insertion is accomplished by ligation,
although any suitable coupling mechanism can also be used. Thus, the only
requirement for nucleic acid molecules to be used in the disclosed method is
15 that they can be coupled to the ends of a double-stranded nucleic acid
25 molecule (that is, the linear vector). Single-stranded nucleic acid molecules,
such as RNA, can be used by converting the molecule to be double-stranded.
In the case of RNA molecules, this can be accomplished, for example, by
30 producing a cDNA molecule of the RNA. Numerous methods are known for
20 preparing and inserting nucleic acid molecules into vectors and any of these
can be used to prepare nucleic acid molecules for use in the disclosed method
(see, for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory*
35 *Manual*, 2nd Edition (Cold Spring Harbor Laboratory Press, Cold Spring
Harbor, N.Y., 1989)). Preferably, the nucleic acid molecule is prepared by
25 generating sticky ends to facilitate insertion in the linear vector. This can be
accomplished, for example by cleaving a nucleic acid molecule of interest, or
40 a nucleic acid sample, with a restriction enzyme, or by adding linkers to the
ends of nucleic acid molecules of interest that have, or can be processed to
have sticky ends. One or both of the ends of the nucleic acid molecule can
45 also be left blunt ended. The two ends of nucleic acid molecules to be used
30 in the disclosed method can also be made different to allow directional
insertion. For example, the two ends can have different sticky ends, or have
one sticky end and one blunt end.

B. Linear Vectors

Linear vectors for use in the disclosed method are double-stranded nucleic acid molecules that can be circularized when its ends are coupled to a nucleic acid molecule. The characteristics of the linear vector are limited only by the requirements for the circular vector that results upon insertion of the nucleic acid molecule. Thus, the linear vector is designed such that, when the nucleic acid molecule is inserted and the linear vector is circularized, the resulting circular vector has one continuous, circular strand and one discontinuous strand.

The use of the term vector is not meant to indicate that the linear vector is required to have any characteristics beyond these, such as promoters, selectable markers, origins of replication, and other features present on traditional vectors for cloning in cells. However, the linear vector may contain these or any other features that do not interfere with the disclosed method. Such additional characteristics may be useful, for example, to allow transfer of the vector to cells to obtain expression, or to allow in vitro expression. Thus, the linear vector can be as simple as a short linker that facilitates circularization of a nucleic acid molecule of interest.

The linear vector is a double-stranded nucleic acid molecule where one of the strands will become part of the continuous strand of the circular vector and the other strand will become part of the discontinuous strand of the circular vector. For identification, the strand of the linear vector which will become part of the continuous strand of the circular vector is referred to as the first strand of the linear vector, and the strand of the linear vector which will become part of the discontinuous strand of the circular vector is referred to as the second strand of the linear vector. The first strand of a linear vector includes a sequence complementary to a rolling circle replication primer. This sequence is referred to as the primer complement portion of the linear vector. This facilitates amplification of the circular vector formed from the linear vector by rolling circle replication primed by the rolling circle replication primer. A separate primer complement portion is not required if the second strand of the circular vector is to serve as the rolling circle replication primer. It is preferred that the primer complement

5 portion of the linear vector is near the 5' end of the first strand of the linear vector.

10 The production of a circular vector in the disclosed method is facilitated by giving the first strand and the second strand of the linear vector 5 different characteristics. Although the linear vector is referred to as having two strands, each of these "strands" may be made up of more than one linear nucleic acid strands. That is, the first strand of the linear nucleic acid molecule may be made up of multiple nucleic acid molecules lying end to 15 end and hybridized to the second strand. Similarly, the second strand of the linear nucleic acid molecule may be made up of multiple strands hybridized to the first strand. The use of the terms first "strand" and second "strand" are 20 used as a convenience to refer to all of the physical nucleic acid strands that make up one side of the linear vector. The relationship of the physical strands in the linear vector to the collective first and second strands can be 25 seen in Figure 1. The first strand of the linear vector is preferably composed of one strand. The second strand is preferably composed of more than one strand, and most preferably composed of two strands.

30 All of the ends present in the first strand of the linear vector, including internal ends, if present, should be ligatable. Ligatable ends are 20 ends that can be ligated to compatible ends by ligase, or which can otherwise be coupled to compatible ends. Preferred ligatable ends are nucleotides having a 3' hydroxyl or a 5' phosphate. Internal ends are ligatable only if 35 compatible ends are adjacent. For example, a nick with a 3' hydroxyl on one end and a 5' phosphate on the other end is a ligatable nick and the ends are 25 ligatable. Nick has its usual meaning. Specifically, a nick is a break in a strand hybridized to another strand where there are no unpaired nucleotides in the other strand opposite the nick. 40

45 To result in a discontinuous second strand in the circular vector, the second strand of the linear vector should contain at least one non-ligatable 30 end or at least one gap or overlap. Non-ligatable ends are ends that cannot be ligated to compatible ends by ligase, or which cannot otherwise be coupled to compatible ends. Preferred non-ligatable ends are nucleotides having a blocking group at the 3' or 5' position. For example, the second strand of the 50

linear vector can include a 3'-terminal or 5'-terminal biotin residue (either at the end of a continuous second strand or at a nick in a discontinuous second strand). This residue renders the terminus non-ligatable, causing all vectors to contain a nick after cloning of inserts by ligation. This biotin residue can then be used as a handle to remove the second strand of the circularized vector, generating single-stranded circles for amplification. Thus, the biotin is both a blocking group and an affinity tag.

Internal ends are also non-ligatable if, for example, compatible ends are not adjacent. For example, a nick with a 3' hydroxyl on both ends is an unligatable nick and the ends are unligatable. A nick with a blocking group on one of the ends is also an unligatable nick and the end with the blocking group is an unligatable end. Both gaps and overlaps are not ligatable even if the ends would otherwise be compatible since the ends are not close enough to be coupled. Gap has its usual meaning. Specifically, a gap is a break in a strand hybridized to another strand where there is at least one nucleotide on the other strand opposite the gap that is unpaired. A gap can also occur at the end of the linear vector in that a nucleic acid molecule, when hybridized to sticky ends of the linear vector, can fail to extend to the end of one of the strands of the linear vector.

An overlap occurs where the adjacent ends of two strands hybridized adjacent to each other on another strand extend beyond the region of hybridization. A preferred form of overlap is where the two overlapping strands hybridize to each other in the overlapping region. This type of overlap in a linear vector produces a Y shaped molecule such as the one illustrated in Figure 1 and in Figure 3.

The second strand of the linear vector may contain multiple nicks, gaps, and overlaps in any combination. Any number of such nicks may be ligatable or non-ligatable. All that is required is at least one feature that prevents the second strand of the circular vector from being continuous following insertion of the nucleic acid molecule. For this purpose, a single non-ligatable end or other non-ligatable feature is all that is required. The first strand of the linear vector contains multiple nicks so long as they are all

ligatable; that is, so long as the first strand of the circular vector will be continuous following insertion of the nucleic acid molecule.

The second strand of the linear vector can also contain one or more affinity tags to facilitate separation of the first and second strands of the circular vector formed from the linear vector. It is preferred that linear vectors include either pre-formed sticky ends or one or more restriction enzymes sites near the ends of the linear vector to facilitate insertion of nucleic acid molecules into the vectors. Multiple cloning sites (MCS) are particularly preferred. Such MCSs facilitate both insertion of a nucleic acid molecule of interest into linear vectors and removal of the nucleic acid molecule from the amplified nucleic acid. It is preferred that the ends of the linear vector, when ready for ligation, do not contain compatible ends that can be ligated. This will prevent the circularization of linear vectors in the absence of insertion of a nucleic acid molecule.

C. Circular Vectors

A circular vector is a double-stranded circular nucleic acid molecule that is a combination of a linear vector and one or more inserted nucleic acid molecules. One of the strands of the circular vector, termed the first strand, is continuous. That is, the first strand of the circular vector is a closed circular nucleic acid strand. The other strand of the circular vector, termed the second strand, is discontinuous. That is, the second strand of the circular vector is not a closed circular nucleic acid strand. The second strand can include, for example, nicks, gaps, and overlaps. The discontinuity of the second strand allows the separation of the first and second strands following denaturation. The second strand of the circular vector can also contain one or more affinity tags to facilitate separation of the first and second strands of the circular vector.

The first strand of a circular vector includes a sequence complementary to a rolling circle replication primer. This sequence is referred to as the primer complement portion of the circular vector. This facilitates amplification of the circular vector by rolling circle replication primed by the rolling circle replication primer. A separate primer

complement portion is not required if the second strand of the circular vector is to serve as the rolling circle replication primer.

D. Affinity Tags

An affinity tag is a molecule that interacts specifically with a particular molecule or moiety. The molecule or moiety that interacts specifically with an affinity tag is referred to herein as an affinity target. Together, an affinity tag and affinity target make up a binding pair. Either member of a binding pair can be used as an affinity tag and either member can be used as an affinity target. An affinity tag is the member of the binding pair coupled to the linear or circular vector. A preferred binding pair is biotin and streptavidin. It is to be understood that the term affinity target refers to both separate molecules and to portions of molecules, such as an epitope of a protein, that interacts specifically with an affinity tag. Antibodies, either member of a receptor/ligand pair, and other molecules with specific binding affinities are examples of affinity tags, useful as the affinity portion of a reporter binding molecule. By coupling an affinity tag to the second strand of a linear vector, binding of the affinity tag to its affinity target allows separation of the first and second strands of the circular vector. An affinity tag that interacts specifically with a particular affinity target is said to be specific for that affinity target. For example, an affinity tag which is an antibody that binds to a particular antigen is said to be specific for that antigen. The antigen is the affinity target. Complementary nucleotide sequences can be used as binding pairs. An example of this is illustrated with the immobilization of Y shaped circular vector in Figure 3.

E. Affinity Substrates

Affinity substrates are solid-state substrates or supports to which affinity targets have been coupled. Generally, an affinity substrate is used to facilitate separation of first and second strands of circular vectors by immobilizing the second strands of circular vectors to a solid-state substrate or support via an affinity tag. Solid-state substrates for use in affinity substrates can include any solid material to which affinity targets can be coupled. This includes materials such as acrylamide, cellulose, nitrocellulose, polystyrene, polyethylene vinyl acetate, polypropylene,

polymethacrylate, polyethylene, polyethylene oxide, glass, polysilicates, polycarbonates, teflon, fluorocarbons, nylon, silicon rubber, polyanhydrides, polyglycolic acid, polylactic acid, polyorthoesters, polypropylfumerate, collagen, glycosaminoglycans, polyamino acids, chemical resistant metals, and corrosion resistant metals. Solid-state substrates can have any useful form including thin films or membranes, beads, bottles, dishes, fibers, woven fibers, shaped polymers, particles and microparticles. A preferred form for a solid-state substrate is a bead or surface.

Affinity targets immobilized on a solid-state substrate allow capture of the second strand of circular vectors on a affinity substrate. Such capture provides a convenient means of separating the second strands from the first strands of circular vectors (which are to be amplified).

Methods for immobilizing proteins, such as antibodies, to solid-state substrates are well established. Immobilization can be accomplished by attachment, for example, to aminated surfaces, carboxylated surfaces or hydroxylated surfaces using standard immobilization chemistries. Examples of attachment agents are cyanogen bromide, succinimide, aldehydes, tosyl chloride, avidin-biotin, photocrosslinkable agents, epoxides and maleimides. A preferred attachment agent is glutaraldehyde. These and other attachment agents, as well as methods for their use in attachment, are described in *Protein immobilization: fundamentals and applications*, Richard F. Taylor, ed. (M. Dekker, New York, 1991), Johnstone and Thorpe, *Immunochemistry In Practice* (Blackwell Scientific Publications, Oxford, England, 1987) pages 209-216 and 241-242, and *Immobilized Affinity Ligands*, Craig T.

Hermanson *et al.*, eds. (Academic Press, New York, 1992). Proteins, and other affinity targets having free amino groups, can be attached to a substrate by chemically cross-linking a free amino group on the protein to reactive side groups present within the solid-state substrate. For example, proteins may be chemically cross-linked to a substrate that contains free amino or carboxyl groups using glutaraldehyde or carbodiimides as cross-linker agents. In this method, aqueous solutions containing free proteins are incubated with the solid-state substrate in the presence of glutaraldehyde or carbodiimide. For crosslinking with glutaraldehyde the reactants can be

incubated with 2% glutaraldehyde by volume in a buffered solution such as 0.1 M sodium cacodylate at pH 7.4. Other standard immobilization chemistries are known by those of skill in the art.

Methods for immobilization of oligonucleotides to solid-state substrates are well established. Oligonucleotides, including affinity targets, can be coupled to substrates using established coupling methods. For example, suitable attachment methods are described by Pease *et al.*, *Proc. Natl. Acad. Sci. USA* 91(11):5022-5026 (1994), and Khrapko *et al.*, *Mol Biol (Mosk) (USSR)* 25:718-730 (1991). A method for immobilization of 3'-amine oligonucleotides on casein-coated slides is described by Stimpson *et al.*, *Proc. Natl. Acad. Sci. USA* 92:6379-6383 (1995). A preferred method of attaching oligonucleotides to solid-state substrates is described by Guo *et al.*, *Nucleic Acids Res.* 22:5456-5465 (1994).

F. Rolling Circle Replication Primer

A rolling circle replication primer (RCRP) is an oligonucleotide having sequence complementary to the primer complement portion of the first strand of a circular vector. This sequence is referred to as the complementary portion of the RCRP. The complementary portion of a RCRP and the cognate primer complement portion can have any desired sequence so long as they are complementary to each other. In general, the sequence of the RCRP can be chosen such that it is not significantly complementary to any other portion of the circular vector. The complementary portion of a rolling circle replication primer can be any length that supports specific and stable hybridization between the primer and the primer complement portion. Generally this is 10 to 35 nucleotides long, but is preferably 16 to 20 nucleotides long. A separate rolling circle replication primer is not required if the second strand of the circular vector is to serve as the rolling circle replication primer. In this case, the second strand of the circular vector can be referred to as a rolling circle replication primer.

It is preferred that rolling circle replication primers also contain additional sequence at the 5' end of the RCRP that is not complementary to any part of the circular vector. This sequence is referred to as the non-

complementary portion of the RCRP. The non-complementary portion of the RCRP, if present, serves to facilitate strand displacement during DNA replication. The non-complementary portion of a RCRP may be any length, but is generally 1 to 100 nucleotides long, and preferably 4 to 8 nucleotides long. The rolling circle replication primer may also include modified nucleotides to make it resistant to exonuclease digestion. For example, the primer can have three or four phosphorothioate linkages between nucleotides at the 5' end of the primer. Such nuclease resistant primers allow selective degradation of excess unligated linear vector that might otherwise interfere with hybridization of probes and primers to the amplified nucleic acid. A rolling circle replication primer can be used as the tertiary strand displacement primer in strand displacement cascade amplification.

G. Strand Displacement Primers

Primers used for strand displacement replication are referred to herein as strand displacement primers. One form of strand displacement primer, referred to herein as a secondary strand displacement primer, is an oligonucleotide having sequence matching part of the sequence of the first strand of a circular vector. This sequence is referred to as the matching portion of the strand displacement primer. This matching portion of a secondary strand displacement primer is complementary to sequences in tandem sequence DNA (TS-DNA). The matching portion of a secondary strand displacement primer may be complementary to any sequence in TS-DNA. However, it is preferred that it not be complementary TS-DNA sequence matching either the rolling circle replication primer or a tertiary strand displacement primer, if one is being used. This prevents hybridization of the primers to each other. The matching portion of a strand displacement primer may be complementary to all or a portion of the inserted nucleic acid molecule, although this is not preferred. The matching portion of a strand displacement primer can be any length that supports specific and stable hybridization between the primer and its complement. Generally this is 12 to 35 nucleotides long, but is preferably 18 to 25 nucleotides long. It is preferred that the matching portion of the circular vector is near the 3' end of the first strand of the circular vector.

5 It is preferred that secondary strand displacement primers also
contain additional sequence at their 5' end that does not match any part of the
first strand of the circular vector. This sequence is referred to as the non-
10 matching portion of the strand displacement primer. The non-matching
5 portion of the strand displacement primer, if present, serves to facilitate
strand displacement during DNA replication. The non-matching portion of a
strand displacement primer may be any length, but is generally 1 to 100
15 nucleotides long, and preferably 4 to 8 nucleotides long.

Another form of strand displacement primer, referred to herein as a
10 tertiary strand displacement primer, is an oligonucleotide having sequence
complementary to part of the sequence of the first strand of the circular
vector. This sequence is referred to as the complementary portion of the
20 tertiary strand displacement primer. This complementary portion of the
tertiary strand displacement primer matches sequences in TS-DNA. The
15 complementary portion of a tertiary strand displacement primer may be
complementary to any sequence in the first strand of the circular vector.
However, it is preferred that it not be complementary to a sequence matching
the strand displacement primer. This prevents hybridization of the primers to
30 each other. The complementary portion of a tertiary strand displacement
20 primer can be any length that supports specific and stable hybridization
between the primer and its complement. Generally this is 12 to 35
nucleotides long, but is preferably 18 to 25 nucleotides long. It is preferred
35 that tertiary strand displacement primers also contain additional sequence at
their 5' end that is not complementary to any part of the first strand of the
25 circular vector. This sequence is referred to as the non-complementary
portion of the tertiary strand displacement primer. The non-complementary
40 portion of the tertiary strand displacement primer, if present, serves to
facilitate strand displacement during DNA replication. The non-
complementary portion of a tertiary strand displacement primer may be any
45 30 length, but is generally 1 to 100 nucleotides long, and preferably 4 to 8
nucleotides long. A rolling circle replication primer is a preferred form of
tertiary strand displacement primer. It is preferred that the complementary

5 portion of the circular vector is near the 5' end of the first strand of the
circular vector.

Strand displacement primers may also include modified nucleotides
to make them resistant to exonuclease digestion. For example, the primer
10 5 can have three or four phosphorothioate linkages between nucleotides at the
5' end of the primer. Such nuclease resistant primers allow selective
degradation of excess unligated linear vectors that might otherwise interfere
15 with hybridization of probes and primers to the amplified nucleic acid.
Strand displacement primers can be used for strand displacement replication
10 and strand displacement cascade amplification, both described below.

H. Synthesis of Oligonucleotides

20 Linear vectors, rolling circle replication primers, strand displacement
primers, and any other oligonucleotides can be synthesized using established
oligonucleotide synthesis methods. Methods to produce or synthesize
15 oligonucleotides are well known in the art. Such methods can range from
25 standard enzymatic digestion followed by nucleotide fragment isolation (see
for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*,
2nd Edition (Cold Spring Harbor Laboratory Press, Cold Spring Harbor,
30 N.Y., 1989) Chapters 5, 6) to purely synthetic methods, for example, by the
20 cyanoethyl phosphoramidite method using a Milligen or Beckman System
1Plus DNA synthesizer (for example, Model 8700 automated synthesizer of
Milligen-Bioscience, Burlington, MA or ABI Model 380B). Synthetic
35 methods useful for making oligonucleotides are also described by Ikuta *et al.*,
Ann. Rev. Biochem. 53:323-356 (1984), (phosphotriester and phosphite-
25 triester methods), and Narang *et al.*, *Methods Enzymol.* 65:610-620 (1980),
(phosphotriester method). Protein nucleic acid molecules can be made using
40 known methods such as those described by Nielsen *et al.*, *Bioconjug. Chem.*
5:3-7 (1994).

Many of the oligonucleotides described herein are designed to be
45 30 complementary to certain portions of other oligonucleotides or nucleic acids
such that stable hybrids can be formed between them. The stability of these
hybrids can be calculated using known methods such as those described in
50 Lesnick and Freier, *Biochemistry* 34:10807-10815 (1995), McGraw *et al.*,

Biotechniques 8:674-678 (1990), and Rychlik *et al.*, *Nucleic Acids Res.* 18:6409-6412 (1990).

I. DNA ligases

Any DNA ligase is suitable for use in the disclosed method.

- Preferred ligases are those that preferentially form phosphodiester bonds at nicks in double-stranded DNA. That is, ligases that fail to ligate the free ends of single-stranded DNA at a significant rate are preferred.
- Thermostable ligases are especially preferred. Many suitable ligases are known, such as T4 DNA ligase (Davis *et al.*, *Advanced Bacterial Genetics - A Manual for Genetic Engineering* (Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1980)), *E. coli* DNA ligase (Panasenko *et al.*, *J. Biol. Chem.* 253:4590-4592 (1978)), AMPLIGASE® (Kalin *et al.*, *Mutat. Res.*, 283(2):119-123 (1992); Winn-Deen *et al.*, *Mol Cell Probes* (England) 7(3):179-186 (1993)), Taq DNA ligase (Barany, *Proc. Natl. Acad. Sci. USA* 88:189-193 (1991)), *Thermus thermophilus* DNA ligase (Abbott Laboratories), *Thermus scotoductus* DNA ligase and *Rhodothermus marinus* DNA ligase (Thorbjarnardottir *et al.*, *Gene* 151:177-180 (1995)). T4 DNA ligase is preferred for ligations involving RNA target sequences due to its ability to ligate DNA ends involved in DNA:RNA hybrids (Hsuih *et al.*, *Quantitative detection of HCV RNA using novel ligation-dependent polymerase chain reaction*, American Association for the Study of Liver Diseases (Chicago, IL, November 3-7, 1995)).

J. DNA polymerases

- DNA polymerases useful in rolling circle replication must perform rolling circle replication of primed single-stranded circles. Such polymerases are referred to herein as rolling circle DNA polymerases. For rolling circle replication, it is preferred that a DNA polymerase be capable of displacing the strand complementary to the template strand, termed strand displacement, and lack a 5' to 3' exonuclease activity. Strand displacement is necessary to result in synthesis of multiple tandem copies of the circular vector. A 5' to 3' exonuclease activity, if present, might result in the destruction of the synthesized strand. It is also preferred that DNA polymerases for use in the disclosed method are highly processive. The

5 suitability of a DNA polymerase for use in the disclosed method can be readily determined by assessing its ability to carry out rolling circle replication. Preferred rolling circle DNA polymerases are bacteriophage ϕ 29 DNA polymerase (U.S. Patent Nos. 5,198,543 and 5,001,050 to Blanco *et*
10 5 *al.*), phage M2 DNA polymerase (Matsumoto *et al.*, *Gene* 84:247 (1989)), phage ϕ PRD1 DNA polymerase (Jung *et al.*, *Proc. Natl. Acad. Sci. USA* 84:8287 (1987)), VENT® DNA polymerase (Kong *et al.*, *J. Biol. Chem.* 268:1965-1975 (1993)), Klenow fragment of DNA polymerase I (Jacobsen *et*
15 *al.*, *Eur. J. Biochem.* 45:623-627 (1974)), T5 DNA polymerase (Chatterjee *et al.*, *Gene* 97:13-19 (1991)), PRD1 DNA polymerase (Zhu and Ito, *Biochim. Biophys. Acta.* 1219:267-276 (1994)), modified T7 DNA polymerase (Tabor and Richardson, *J. Biol. Chem.* 262:15330-15333 (1987); Tabor and Richardson, *J. Biol. Chem.* 264:6447-6458 (1989); Sequenase™ (U.S. Biochemicals)), and T4 DNA polymerase holoenzyme (Kaboord and
20 15 Benkovic, *Curr. Biol.* 5:149-157 (1995)). ϕ 29 DNA polymerase is most preferred. Rolling circle DNA polymerases are also generally useful for strand displacement replication.

Strand displacement can be facilitated through the use of a strand displacement factor, such as helicase. It is considered that any DNA
30 20 polymerase that can perform rolling circle replication in the presence of a strand displacement factor is suitable for use in the disclosed method, even if the DNA polymerase does not perform rolling circle replication in the absence of such a factor. Strand displacement factors useful in RCA include BMRF1 polymerase accessory subunit (Tsurumi *et al.*, *J. Virology* 67(12):7648-7653 (1993)), adenovirus DNA-binding protein (Zijderveld and van der Vliet, *J. Virology* 68(2):1158-1164 (1994)), herpes simplex viral protein ICP8 (Boehmer and Lehman, *J. Virology* 67(2):711-715 (1993); Skaliter and Lehman, *Proc. Natl. Acad. Sci. USA* 91(22):10665-10669 (1994)), single-stranded DNA binding proteins (SSB; Rigler and Romano, *J.*
40 25 *Biol. Chem.* 270:8910-8919 (1995)), and calf thymus helicase (Siegel *et al.*, *J. Biol. Chem.* 267:13629-13635 (1992)).

The ability of a polymerase to carry out rolling circle replication can be determined by using the polymerase in a rolling circle replication assay

5 such as those described in Fire and Xu, *Proc. Natl. Acad. Sci. USA* 92:4641-4645 (1995).

10 It is possible to enhance the specificity of the DNA amplification reactions used in the disclosed method by using a DNA polymerase that is
5 inactive at low temperature, and active only at high temperature. An example of such an enzyme, AmpliTaq Gold, has been described by Moretti *et al.*, *Biotechniques* 25:716-722 (1998). AmpliTaq Gold is inactive until heated during the PCR before thermal cycling. A similar enzyme could be
15 used in the disclosed method. Temperature activation of DNA polymerase can also be achieved using antibodies specific for the polymerase. For example, antibodies specific for Bst large fragment DNA polymerase could
20 be obtained by immunization of mice. Among such antibodies, one could be chosen on the basis of its ability to bind to and inhibit the enzyme at room temperature. The antibody could also be chosen, using known screening
25 procedures, such that upon heating, the inhibition of the DNA polymerase would cease. Combining the antibody with Bst large fragment DNA polymerase would generate an enzyme mixture that is activated upon heating.

30 K. Kits

20 Any combination of the materials useful in the disclosed method can be packaged together as a kit for performing the disclosed method. In particular, linear vectors, rolling circle replication primers, affinity
35 substrates, and strand displacement primers are useful components of such kits. Enzymes necessary for the disclosed method are also preferred
25 components of such kits.

40 L. Tandem Sequence DNA

45 The first strand of the circular vector, when replicated, gives rise to a long DNA molecule containing multiple repeats of sequences complementary to the circular vector. This long DNA molecule is referred to
30 herein as tandem sequences DNA (TS-DNA). TS-DNA contains sequences complementary to the inserted nucleic acid molecule and the primer complement portion. If the tandem sequence DNA is itself replicated by
50 strand displacement amplification, the resulting long DNA molecules

5 containing multiple repeats of sequences matching the circular vector are referred to as secondary tandem sequence DNA. If the secondary tandem sequence DNA is in turn replicated by strand displacement amplification, the resulting long DNA molecules containing multiple repeats of sequences
10 5 complementary to the circular vector are referred to as tertiary tandem sequence DNA.

M. Collected Samples (Library Replica)

15 The usefulness of the disclosed method is increased by producing libraries of clones and saving samples of the clones for later use. Such
10 samples are referred to as collected samples. Collecting samples is analogous to replica plating in cell-based cloning. Samples of amplified nucleic acid can be collected, for example, by transfer with an array of pins (most useful
20 when the nucleic acid is amplified in an array pattern), by transfer into an array, by direct transfer from a spread of amplified nucleic acid on a surface to another surface (this is analogous to colony transfer), and by blotting the
25 amplified nucleic acid onto a membrane (most useful when the nucleic acid is amplified in agarose). Once the samples are collected, they can be further amplified to allow analysis or use of the clones, or to allow another round of replica collection.

II. Method

35 The disclosed method involves inserting nucleic acid molecules of interest into a linear vector to form a circular vector with one continuous strand and one discontinuous strand. The discontinuous strand may include
25 an affinity tag which, by interaction with an affinity substrate, can facilitate separation of the continuous strand from the discontinuous strand. The continuous strand of the circular vector is amplified by rolling circle replication to form tandem sequence DNA. Rolling circle replication is primed by a rolling circle replication primer complementary to a sequence in
40 the continuous strand. The tandem sequence DNA itself may be amplified by strand displacement replication, to form secondary tandem sequence DNA, using strand displacement primers (complementary to a sequence in the tandem sequence DNA). The secondary tandem sequence DNA may
45 50

5 also be amplified by strand displacement replication, to form tertiary tandem
sequence DNA, using strand displacement primers or the rolling circle
replication primers (complementary to a sequence in the secondary tandem
sequence DNA). The amplified DNA can be sequenced, probed, subjected
10 5 to restriction analysis, subcloned, transcribed, subjected to hybridization or
denaturation analysis, further amplified, or stored.

Multiple amplification reactions can be carried out in parallel,
preferably in arrays or as spreads of diluted vectors on surfaces or embedded
15 in agarose. The resulting "colonies" of amplified DNA represent molecular
clones of the progenitor circular vectors with an inserted nucleic acid
molecule. Collectively, such colonies form a library of cloned nucleic acid
20 molecules that can be replica plated or arrayed, stored, and screened. These
procedures are described in detail below.

When using the disclosed method to produce a library of cDNA
15 molecules, or to analyze mRNA in a sample via cDNA, the cDNA
preparations used for cloning in the disclosed vectors can be prepared using
methods that reduce the over-representation of cDNA that corresponds to
highly abundant messenger RNA. Libraries made using such methods are
30 called normalized libraries (Bonaldo *et al.*, *Genome Res* 6:791-806 (1996)).
20 The use of normalized libraries reduces the number of clones that must be
screened to find a sequence of interest.

A. Ligation

35 Ligation of nucleic acid molecules into linear vectors can be
accomplished using any suitable conditions. Techniques for insertion of
nucleic acid molecules into vectors in general are well established and can be
25 used with the disclosed linear vectors. Suitable ligases for the ligation
operation are described above. Ligation reactions can involve a single type
of linear vector and a single type of nucleic acid molecule to be inserted, a
single type of linear vector and multiple different types of nucleic acid
45 30 molecules to be inserted, multiple types of linear vector and a single type of
nucleic acid molecule to be inserted, or multiple types of linear vectors and
multiple types of nucleic acid molecules to be inserted. For general cloning
and production of nucleic acid libraries it is preferred that a single type of
50

5 linear vector and multiple different types of nucleic acid molecules to be
inserted be used. For subcloning of specific nucleic acid fragments it is
preferred that a single type of linear vector and a single type of nucleic acid
molecule to be inserted. Ligation conditions are generally known. Most
10 5 ligases require Mg^{++} . There are two main types of ligases, those that are
ATP-dependent and those that are NAD-dependent. ATP or NAD,
depending on the type of ligase, should be present during ligation.

15 Ligation of compatible ends of nucleic acid molecules and vectors
can be facilitated through the use of blunt ends or sticky ends as is known in
the field of molecular cloning. Both blunt ends and sticky ends can be
10 produced by digestion of the nucleic acid molecules and the linear vectors
with appropriate restriction enzymes, by ligation of appropriate linkers to the
ends of the nucleic acid molecules and the linear vectors, or both. In the case
of linear vectors, appropriate ends can be formed directly by the structure of
15 the linear vector without the need for restriction enzyme digestion of linker
ligation. In the case of the nucleic acid molecules to be inserted, appropriate
ends can be appended to the ends during preparation of the nucleic acid
molecule. For example, appropriate ends can be incorporated into cDNA by
20 using primer having appropriate sequences during cDNA synthesis or by
adding a nucleotide tail to the cDNA.

B. Amplification

35 The circular vectors formed by ligation of linear vectors and nucleic
acid molecules of interest serve as substrates for a rolling circle replication.
This reaction requires the addition of two reagents: (a) a rolling circle
25 replication primer, which is complementary to the primer complement
portion of the first strand of the circular vector, and (b) a rolling circle DNA
40 polymerase. The DNA polymerase catalyzes primer extension and strand
displacement in a processive rolling circle polymerization reaction that
proceeds as long as desired, generating a molecule of up to 100,000
45 30 nucleotides or larger that contains up to approximately 25 tandem copies of a
sequence complementary to a 4000 bp circular vector. This tandem
sequence DNA (TS-DNA) consists of alternating vector sequence and insert

sequence. As an alternative, the second strand of the circular vector can serve as the rolling circle replication primer.

During rolling circle replication one may additionally include radioactive, or modified nucleotides such as bromodeoxyuridine triphosphate, in order to label the DNA generated in the reaction. Alternatively, one may include suitable precursors that provide a binding moiety such as biotinylated nucleotides (Langer *et al.*, *Proc. Natl. Acad. Sci. USA* 78:6633 (1981)).

Strand displacement replication is a way to amplify TS-DNA. Strand displacement replication is accomplished by hybridizing strand displacement primers to TS-DNA and allowing a DNA polymerase to synthesize DNA from these primed sites. The product of strand displacement replication is referred to as secondary tandem sequence DNA or TS-DNA-2. Strand displacement replication can be accomplished by performing rolling circle replication to produce TS-DNA, and then mixing strand displacement primer with the TS-DNA and incubating to replicate the tandem sequence DNA. The strand displacement primer is complementary to a part of the circular vector used to generate TS-DNA as described earlier. It is preferred that the strand displacement primer is not complementary to the rolling circle replication primer, or to a tertiary strand displacement primer, if used.

Strand displacement replication can also be carried out simultaneously with rolling circle replication. This is accomplished by mixing strand displacement primer with the circular vector and rolling circle replication primer prior to incubating the mixture for rolling circle replication. For simultaneous rolling circle replication and strand displacement replication, it is preferred that the rolling circle DNA polymerase be used for both replications. This allows optimum conditions to be used and results in displacement of other strands being synthesized downstream. Generally, strand displacement replication can be performed by, simultaneous with or following rolling circle replication, mixing a strand displacement primer with the TS-DNA and incubating to replicate the tandem sequence DNA to result in the formation of secondary tandem sequence DNA.

5 To optimize the efficiency of strand displacement replication, it is
preferred that a sufficient concentration of strand displacement primer be
used to obtain sufficiently rapid priming of the growing TS-DNA strand to
out-compete any remaining unligated linear vectors that might be present for
10 5 binding to TS-DNA. In general, this is accomplished when the strand
displacement primer is in very large excess compared to the concentration of
single-stranded sites for hybridization of the strand displacement primer on
TS-DNA. Optimization of the concentration of strand displacement primer
15 can be aided by analysis of hybridization kinetics using methods such as
those described by Young and Anderson, "Quantitative analysis of solution
hybridization" in *Nucleic Acid Hybridization: A Practical Approach* (IRL
Press, 1985) pages 47-71. Alternatively, the efficiency of strand
20 displacement replication can be improved by the removal of unligated linear
vectors prior to amplification of the TS-DNA. In strand displacement
15 replication, it is preferred that the concentration of strand displacement
primer generally be from 500 nM to 5000 nM, and most preferably from 700
25 nM to 1000 nM.

30 As a strand displacement primer is elongated, the DNA polymerase
will run into the 5' end of the next hybridized strand displacement molecule
20 and will displace its 5' end. In this fashion a tandem queue of elongating
DNA polymerases is formed on the TS-DNA template. As long as the
rolling circle reaction continues, new strand displacement primers and new
35 DNA polymerases are added to TS-DNA at the growing end of the rolling
circle.

25 When strand displacement replication is carried out in the presence of
a tertiary strand displacement primer, an exponential amplification of TS-
40 DNA sequences takes place. This special and preferred mode of strand
displacement replication is referred to as strand displacement cascade
amplification (SDCA). In SDCA, a strand displacement primer primes
45 30 replication of TS-DNA to form TS-DNA-2, as described above. The tertiary
strand displacement primer can then hybridize to, and prime replication of,
TS-DNA-2 to form TS-DNA-3 (tertiary tandem sequence DNA). Strand
50 displacement of TS-DNA-3 by the adjacent, growing TS-DNA-3 strands

5 makes TS-DNA-3 available for hybridization with secondary strand
displacement primer. This results in another round of replication resulting in
TS-DNA-4 (which is equivalent to TS-DNA-2). TS-DNA-4, in turn,
10 becomes a template for DNA replication primed by tertiary strand
5 displacement primer. The cascade continues this manner until the reaction
stops or reagents become limiting. This reaction amplifies DNA at an almost
exponential rate, although kinetics are not truly exponential because there are
15 stochastically distributed priming failures, as well as steric hindrance events
related to the large size of the DNA network produced during the reaction.

10 In a preferred mode of SDCA, the rolling circle replication primer
serves as the tertiary strand displacement primer, thus eliminating the need
20 for a separate primer. For this mode, the rolling circle replication primer
should be used at a concentration sufficiently high to obtain rapid priming on
the growing TS-DNA-2 strands. To optimize the efficiency of SDCA, it is
25 preferred that a sufficient concentration of secondary strand displacement
primer and tertiary strand displacement primer be used to obtain sufficiently
rapid priming of the growing TS-DNA strand to out-compete TS-DNA for
binding to its complementary TS-DNA, and, in the case of secondary strand
30 displacement primer, to out-compete any remaining unligated linear vector
20 that might be present for binding to TS-DNA. In general, this is
accomplished when the secondary strand displacement primer and tertiary
strand displacement primer are both in very large excess compared to the
35 concentration of single-stranded sites for hybridization of the strand
displacement primers on TS-DNA. For example, it is preferred that the
25 secondary strand displacement primer is in excess compared to the
concentration of single-stranded secondary strand displacement primer
40 complement sites on TS-DNA, TS-DNA-3, TS-DNA-5, and so on. In the
case of tertiary strand displacement primer, it is preferred that the tertiary
strand displacement primer is in excess compared to the concentration of
45 30 single-stranded tertiary strand displacement primer complement sites on TS-
DNA-2, TS-DNA-4, TS-DNA-6, and so on. Such an excess generally results
in a primer hybridizing to its complement in TS-DNA before amplified
complementary TS-DNA can hybridize. Optimization of primer
50

5 concentrations can be aided by analysis of hybridization kinetics (Young and
Anderson). In a strand displacement cascade amplification, it is preferred
that the concentration of both secondary and tertiary strand displacement
primers generally be from 500 nM to 5000 nM, and most preferably from
10 5 700 nM to 1000 nM.

As in the case of secondary strand displacement primers, if the
concentration of DNA polymerase is sufficiently high, the polymerase will
initiate DNA synthesis at each available 3' terminus on the hybridized
15 tertiary strand displacement primers, and these elongating TS-DNA-3
10 molecules will block any hybridization by TS-DNA-2. As a tertiary strand
displacement primer is elongated to form TS-DNA-3, the DNA polymerase
will run into the 5' end of the next hybridized tertiary strand displacement
20 primer molecule and will displace its 5' end. In this fashion a tandem queue
of elongating DNA polymerases is formed on the TS-DNA-2 template. As
15 long as the reaction continues, new rolling circle replication primers and new
25 DNA polymerases are added to TS-DNA-2 at the growing ends of TS-DNA-
2. This hybridization/replication/strand displacement cycle is repeated with
hybridization of secondary strand displacement primers on the growing TS-
DNA-3.

20 Generally, strand displacement cascade amplification can be
performed by, simultaneous with, or following, rolling circle replication,
mixing a secondary strand displacement primer and a tertiary strand
35 displacement primer with the TS-DNA and incubating to replicate the
tandem sequence DNA -- where replication of the tandem sequence DNA
25 results in the formation of secondary tandem sequence DNA and where
replication of the secondary tandem sequence DNA results in formation of
40 tertiary tandem sequence DNA (TS-DNA-3).

Strand displacement replication can also be carried out sequentially.
Following a first round of strand displacement replication, a tertiary strand
45 30 displacement primer can be mixed with the TS-DNA and TS-DNA-2 and
incubated to replicate the secondary tandem sequence DNA, where
replication of the secondary tandem sequence DNA results in formation of
50 tertiary tandem sequence DNA (TS-DNA-3). This round of strand

displacement replication can be referred to as tertiary strand displacement replication. However, all rounds of strand displacement replication following rolling circle replication can also be referred to collectively as strand displacement replication.

A modified form of strand displacement replication results in amplification of TS-DNA and is referred to as opposite strand amplification (OSA). OSA is the same as strand displacement replication except that a special form of rolling circle replication primer is used that prevents it from hybridizing to TS-DNA-2. This can be accomplished in a number of ways.

For example, the rolling circle replication primer can have an affinity tag coupled to its non-complementary portion allowing the rolling circle replication primer to be removed prior to strand displacement replication. Alternatively, remaining rolling circle replication primer can be crippled following initiation of rolling circle replication. One preferred form of rolling circle replication primer for use in OSA is designed to form a hairpin that contains a stem of perfectly base-paired nucleotides. The stem can contain 5 to 12 base pairs, most preferably 6 to 9 base pairs. Such a hairpin-forming rolling circle replication primer is a poor primer at lower temperature (less than 40°C) because the hairpin structure prevents it from hybridizing to complementary sequences. The stem should involve a sufficient number of nucleotides in the complementary portion of the rolling circle replication primer to interfere with hybridization of the primer to the circular vector. Generally, it is preferred that a stem involve 5 to 24 nucleotides, and most preferably 6 to 18 nucleotides, of the complementary portion of a rolling circle replication primer. A rolling circle replication primer where half of the stem involves nucleotides in the complementary portion of the rolling circle replication primer and the other half of the stem involves nucleotides in the non-complementary portion of the rolling circle replication primer is most preferred. Such an arrangement eliminates the need for self-complementary regions in the circular vector when using a hairpin-forming rolling circle replication primer.

If an excess of tertiary tandem sequence DNA is desired, the secondary strand displacement primer can be crippled in the same manner as

5 is described above for the rolling circle replication primer (the rolling circle
replication primer and tertiary strand displacement primer should not be
crippled in this case). The reaction at the higher, permissive temperature
should be carried out long enough to produce a reasonable amount of
10 5 secondary tandem sequence DNA to serve as a template for tertiary sequence
DNA. When the temperature is shifted, the secondary strand displacement
primer can no longer prime synthesis and the synthesis of tertiary tandem
sequence DNA soon outstrips the amount of secondary tandem sequence
15 DNA. Of course tandem sequence DNA will continue to be produced by
20 rolling circle replication throughout the reaction (since the rolling circle
replication primer is not crippled).

20 When starting the rolling circle replication reaction, secondary strand
displacement primer and rolling circle replication primer are added to the
reaction mixture, and the solution is incubated briefly at a temperature
15 sufficient to disrupt the hairpin structure of the rolling circle replication
25 primer but to still allow hybridization to the primer complement portion of
the circular vector (typically greater than 50°C). This incubation permits the
rolling circle replication primer to hybridize to the primer complement
portion of the circular vector. The solution is then brought to the proper
30 temperature for rolling circle replication, and the rolling circle DNA
polymerase is added. As the rolling circle reaction proceeds, TS-DNA is
generated, and as the TS-DNA grows in length, the secondary strand
35 displacement primer rapidly initiates DNA synthesis with multiple strand
displacement reactions on TS-DNA. These reactions generate TS-DNA-2,
25 which is complementary to the TS-DNA. While TS-DNA-2 contains
sequences complementary to the rolling circle replication primer, the primer
is not able to hybridize nor prime efficiently at the reaction temperature due
40 to its hairpin structure at this temperature. Thus, there is no further priming
by the rolling circle replication primer and the only products generated are
45 30 TS-DNA and TS-DNA-2. The reaction comes to a halt as rolling circle
amplification stops and TS-DNA becomes completely double-stranded. In
the course of the reaction, an excess of single-stranded TS-DNA-2 is
50 generated.

Another form of rolling circle replication primer useful in OSA is a chimera of DNA and RNA. In this embodiment, the rolling circle primer has deoxyribonucleotides at its 3' end and ribonucleotides in the remainder of the primer. It is preferred that the rolling circle replication primer have five or six deoxyribonucleotides at its 3' end. By making part of the rolling circle replication primer with ribonucleotide, the primer can be selectively degraded by RNase H when it is hybridized to DNA. Such hybrids form during OSA as TS-DNA-2 is synthesized. The deoxyribonucleotides at the 3' end allow the rolling circle DNA polymerase to initiate rolling circle replication. RNase H can then be added to the OSA reaction to prevent priming of TS-DNA-2 replication.

Unligated linear vectors may be removed prior to rolling circle replication to eliminate competition between unligated linear vectors and the secondary strand displacement primer for hybridization to TS-DNA.

Alternatively, the concentration of the secondary strand displacement primer can be made sufficiently high so that it out-competes unligated linear vector for hybridization to TS-DNA. This allows strand displacement replication to be performed without removal of unligated linear vectors.

C. Separation

Once the linear vector and nucleic acid molecule are circularized to form a circular vector with one continuous strand and one discontinuous strand, it is preferred that the continuous strand of the circular vector (a single-stranded, closed circular nucleic acid molecule susceptible to rolling circle replication) is separated from the discontinuous strand of the circular vector. Separation allows rolling circle replication to proceed more efficiently.

It is preferred that the two strands of the circular vector are separated by immobilizing the discontinuous (second) strand of the circular vector and then denaturing and washing away the continuous (first) strand of the circular vector. This can be accomplished, for example, by including an affinity tag in the second strand of the circular vector which can then be bound to an immobilized affinity target, thus immobilizing the second strand

5 of the circular vector. For this purpose, use of biotin as an affinity tag and streptavidin as an affinity target is preferred.

Complementary oligonucleotides can also be used as binding pairs for separating the first and second strands of the circular vector. An
10 5 oligonucleotide affinity tag that is a part of or coupled to the second strand of the circular vector is hybridized to an immobilized complementary oligonucleotide. The oligonucleotide affinity tag is preferably an
15 unhybridized tail in an overlap in the second strand of the circular vector (see Figure 3). It is preferred that the oligonucleotide affinity tag be ligated or
20 10 otherwise coupled to a solid-state substrate or support to keep the second strand immobilized during denaturation. This can be accomplished, for example, by using a circular vector with a staggered tail in the second strand
25 and an immobilized oligonucleotide that can hybridize to the longer strand of the tail such that the end of the immobilized oligonucleotide can be ligated to
30 15 the shorter strand of the staggered tail. An example of such an arrangement is shown in Figure 3.

The first and second strands of the circular vector can be denatured using any suitable means. Preferred conditions for denaturation include the
30 use of heat, alkaline conditions, chaotropic conditions, and combinations. These and other means of nucleic acid denaturation are known and can be
35 20 used in the disclosed method. Washing and collection of the first strand is performed during or after denaturation.

As an example, the first and second strands of circular can be separated using a two-step procedure. First, the circular vector, which
40 25 contains a biotin residue on the second strand, is bound to beads containing streptavidin, in order to bind the vector via the biotin. The beads are then washed with formamide at mildly alkaline pH. Under appropriate
45 conditions, the circular vector, which contains an unligated nick site by design, separates into two DNA molecules. Thus, alkaline denaturation
50 30 releases free single-stranded circles from the beads. The single-stranded circular molecules (that is, the first strands of the circular vector) are then further purified by gel filtration or ion exchange (Mono-Q 5/5) chromatography in the presence of an alkaline buffer (15 mM NaOH). This

5 purification step will remove small circles, and small linear vector molecules
that contaminate the circular vectors with inserts.

D. Removing Linear Nucleic Acids

Unligated linear nucleic acids, including unligated linear vector, can
10 5 be removed prior to rolling circle replication. In addition to methods
described elsewhere herein, the gene 6 exonuclease of phage T7 provides a
useful tool for the elimination of linear nucleic acids that might bind to the
15 TS-DNA. This exonuclease digests DNA starting from the 5' end of a
double-stranded structure. It has been used successfully for the generation of
20 single-stranded DNA after PCR amplification (Holloway *et al.*, *Nucleic
Acids Res.* 21:3905-3906 (1993); Nikiforov *et al.*, *PCR Methods and
Applications* 3:285-291(1994)). This enzyme can be added after ligation,
together with the rolling circle DNA polymerase. To protect TS-DNA from
25 degradation, the rolling circle replication primer can contain 3 or 4
phosphorothioate linkages at the 5' end, to make this molecule resistant to
the exonuclease (Nikiforov *et al.* (1994)). The exonuclease will degrade
unprotected linear molecules as they become associated with the rolling
circle DNA product.

E. Dilution and Division

30 The disclosed method is particularly useful for creation of a library of
cloned nucleic acid molecules. For this purpose it is useful to dilute and
divide ligated vectors containing inserts to separate individual circular
35 vectors and allow production of clonal "colonies" of nucleic acid amplified
from a single circular vector. It is preferred that a solution containing
25 circular vectors is diluted sufficiently such that amplification reactions
contain, on average, a single circular vector. This can be accomplished, for
40 example, by making a range of dilutions, dividing the diluted vector
solutions among reactions, and performing amplification. The dilution is
considered optimal when about 33% of the reactions produce amplified
45 nucleic acid. This follows from well known distribution statistics.

The division of the diluted circular vectors can be accomplished in
several ways. For example, the diluted solution can be spotted as an array on
50 a surface. The diluted solution can also be spread on a surface with the

5 circular vectors becoming separated. The diluted solution can also be mixed
with agarose and spread on a surface. A preferred way to divide the diluted
10 circular vectors is to use a microarray of very small liquid droplets on a glass
surface. The micro-droplet arrays contain the diluted circular vectors, and
5 enable the generation of DNA clones in very small compartments, without
the need for physical barriers such as tubes or wells.

15 One useful way to combine division of the ligation reaction with
amplification is to spread the ligation reaction (diluted or not as appropriate)
on a surface and then to spot amplification reaction components (that is,
10 buffers, reagents, polymerase) on the surface in an array. Techniques for
spotting are described, for example, in U.S. Patent No. 5,807,522 to Brown
20 et al. The method here differs, however, in that it is the reaction
components, rather than individual samples, that are spotted. All of the
spotted material is identical. The amplification reagents will then cause
25 amplification of whatever nucleic acid is present at the location of each spot
of reagents. The nucleic acid on the surface comes from the spread of the
ligation reaction.

30 As an example of spreading, the ligated DNA can be diluted serially
to obtain concentrations in the range of several million molecules per
20 milliliter. Approximately 22 μ l of this DNA solution (containing
approximately 30,000 to 120,000 DNA molecules) is placed on a cover slip,
35 and covered with a polylysine-coated microscope slide. The DNA is allowed
to bind the polylysine-covered surface for 30 minutes at 37°C. The slide
then dipped in 0.01% Tween-20, and dried at room temperature. Using an
40 arraying instrument, an array is constructed consisting of 6000 individual
micro-droplets of a solution containing the four dideoxynucleoside
triphosphates, a suitable buffer, and enzymes for amplification. The
45 diameter of the droplets can be approximately 0.150 millimeters. Droplets
preferably are dispensed on the surface of the slide in a controlled humidity
30 atmosphere, in order to maintain a constant droplet volume during
amplification. Alternatively, the diluted circular vectors may be placed on
the glass surface using the arrayer instrument to dispense small volumes that
50 on the average contain a single ligated DNA molecule. It can be calculated

5 that when 33% of the droplets grow molecular colonies, and the rest do not
grow anything, most colonies are likely to be of clonal origin. Adjusting the
initial inoculum density, it should be possible to obtain up to 1500 clonal
10 colonies per 6000-droplet array.

5 As an example of agarose spreading, dilutions of the vectors can be
mixed with a buffer containing melted agarose at 60°C and overlaying the
solution on a petri dish to form a thin agarose layer (0.2 to 2% agarose), such
15 that the concentration of the vectors is in the range of 500 to 5,000 per plate.
The embedded vectors can then be amplified using the disclosed method. At
20 appropriate dilutions, DNA molecular colonies clonally derived from single
vectors will form in the thin film of agarose. The initial density of seed
DNA molecules should be such that the DNA molecular colonies do not
overlap. One useful technique of agarose spreading is describe in U.S.
Patent No. 5,616,478 to Cherverin et al.

25 15 **F. Sample Collection (Replica Plating)**

The usefulness of the disclosed method is increased by producing
libraries of clones and saving samples of the clones for later use. Such
30 samples are referred to as collected samples. Collecting samples is
analogous to replica plating in cell-based cloning. Samples of amplified
20 nucleic acid can be collected, for example, by transfer with an array of pins
(most useful when the nucleic acid is amplified in an array pattern), by
35 transfer into an array, by direct transfer from a spread of amplified nucleic
acid on a surface to another surface (this is analogous to colony transfer), and
by blotting the amplified nucleic acid unto a membrane (most useful when
25 the nucleic acid is amplified in agarose). Once the samples are collected,
40 they can be further amplified to allow analysis or use of the clones, or to
allow another round of replica collection.

Where droplet arrays are used, the molecular colonies (that is, the
45 droplets following amplification) can be replicated by contacting the array
30 with a multi-pin replicator that will bind only a fraction of the volume of the
micro-droplet. Colony replicas may be stored by blotting the replicator on a
membrane such as nitrocellulose of NA-45 (S&S). A preferred way to store
50 replicas is to contact the replicator with a polylysine-coated glass slide,

5 which will permit hybridization or primer extension sequencing with
fluorescent probes. Replicas of reaction droplets on a surface can also be
made by contacting a second surface with the droplets. In general, it is
10 5 preferred that the second surface contact only the droplets and not the first
surface. Replicas of spreads of nucleic acids on a surface likewise can be
made by contacting a second surface with the spread.

Replicas of amplified nucleic acid bound covalently to glass can be
made using any suitable coupling procedure. For example, in order to
15 facilitate a subsequent step of covalent binding to a glass surface, each of the
two strand displacement replication primers used for the strand displacement
20 cascade amplification of circular vectors may be synthesized with a primary
amino group at the 5' end. At the end of the amplification reaction, the glass
slide will contain thousands of liquid droplets harboring DNA clones, and all
the DNA molecules will contain 5'-terminal reactive amino groups. At this
15 point the glass slide can be contacted with another glass slide, leaving an air
gap of less than 1 mm (defined by the thickness of a plastic spacer), in such a
manner that the glass slide on top will contact all of the liquid droplets
without excessive compression. The lower face of this slide (the upper slide)
30 can be derivatized using the methods described by Guo *et al.*, *Nucleic Acids
Research* 22:5456-5465 (1994), Guo *et al.*, *Nature Biotechnology*
15:331-335 (1997), Guo *et al.*, *Nucleic Acids Res* 22:5456-5465 (1994), or
Beier and Hoheisel, *Nucleic Acids Res* 27:1970-1977 (1999), to make the
35 glass surface chemically reactive. The two-slide sandwich is incubated for 1
to 2 hours at 37°C (or as appropriate for the derivative chemistry involved)
25 in order to obtain covalent coupling of the amplified DNA contained in each
droplet to the lower face of the upper slide.

Where agarose has been used to form molecular colonies, analysis of
the colonies can be facilitated by blotting the amplified nucleic acid onto a
membrane or other blotting surface. Many such blotting techniques are
45 30 known and can be used with the disclosed method. For example, the agarose
film can be placed in a vacuum-blotting device that contacts both the bottom
and the top of the agarose film. The nucleic acid on the agarose film can
then be vacuum-blotted to two membranes simultaneously, one placed on top

5 of the agarose and the other below the agarose, to generate two replicas of the molecular colonies on the surface of the membrane.

G. Detecting Amplified Nucleic Acid Molecules

10 The amplified nucleic acid can be used for any purpose for which nucleic acids can be used. For example, the nucleic acid can be sequenced, probed, subjected to restriction analysis, subcloned, transcribed, subjected to hybridization or denaturation analysis, further amplified, or stored. Diagnostic methods, such as sequencing and probing for specific sequences, are preferred. For these purposes, the amplified nucleic acid can be analyzed using standard molecular biology procedures, such restriction enzyme digestion, cloning in a plasmid vector, PCR amplification, which are well known.

15 Libraries of cloned nucleic acids formed by the disclosed method can be screened using any of the methods used for screening conventional libraries. For example, cDNA libraries made using the disclosed method can be analyzed using conventional screens. Libraries can also be used for *in situ* transcription to generate RNA colonies, which can then be analyzed (*in situ* or in replicas) by appropriate screens, such as aptamer screens or ribozyme activity screens. Libraries can also be screened by *in situ* translation on array replicas (see, for example, Saris *et al.*, *Nucleic Acids Res.* 10:4831-4843 (1982)). Libraries can also be screened by *in situ* coupled transcription-translation systems, and subsequent catalytic activity assays for the analysis of mutagenized enzymes.

20 The disclosed method can also be used for serial analysis of gene expression (SAGE), making it more efficient by streamlining the cloning and sequencing into a single process stream. This method involves amplification of cDNA inserted into linear vectors as described herein prior to the SAGE analysis. This means of amplification is useful since PCR amplification of the cDNA prior to cloning, which can skew the abundance of cDNA sequences due to differential amplification, is avoided. The disclosed method insures that sequence tag frequencies in the clone population (which are measured in SAGE) reflect the original frequencies of the cDNAs.

5 The method of Welford *et al.*, *Nucleic Acids Res.* 26(12):3059-3065
(1998), in which thousands of colonies produced using laborious traditional
procedures were analyzed in an array, can be modified to make use of the
disclosed method and thereby become more streamlined and efficient. The
10 5 method could even be automated. Detection of differences between nucleic
acid samples or probe sets can be accomplished by adapting the technique
described by George *et al.*, *Nucleic Acids Research* 27:1517-1523 (1999), to
the disclosed method. George *et al.* describes combination of suppression
15 subtractive hybridization (SSH) and cDNA microarrays for rapid
identification of differentially expressed genes. In this method, a set of
cDNA clones, including inserts amplified by PCR, is arrayed using robotic
printing. The cDNA arrays can then be hybridized with fluorescent labeled
20 probes prepared from RNA obtained from a cell line or tissue of interest.

H. Sequencing Amplified Nucleic Acid Molecules

15 The amplified nucleic acid can be sequenced using any suitable
25 procedure. Many such procedures are known. One preferred form of
sequencing for use with amplified sequences produced with the disclosed
method is nanosequencing or single-nucleotide extension sequencing.
Nanosequencing methods are described below and by Jalanko *et al.*, *Clinical*
30 20 *Chemistry* 38:39-43 (1992); Nikiforov *et al.*, *Nucleic Acids Research*
22:4167-4175 (1994); and Kobayashi *et al.*, *Molecular and Cellular Probes*
9:175-182 (1995).

35 Two forms of primer extension sequencing that can be used with the
disclosed method are described in PCT Application WO 97/20948. One is
25 single nucleotide primer extension sequencing involving interrogation of a
single nucleotide in an amplified target sequence by incorporation of a
40 specific and identifiable nucleotide based on the identity of the interrogated
nucleotide. The other is degenerate probe primer extension sequencing
involving sequential addition of degenerate probes to an interrogation primer
45 30 hybridized to amplified target sequences.

Nanosequencing operations can be performed in batch. For example,
if the slide contains 3000 dots, all 3000 dots are sequenced in a single batch
operation. This can be accomplished by washing the slide with 1% ammonia

after imaging of the first primer extension reaction. This alkaline solution denatures the labeled primer, but the cloned DNA remains on the slide because it is bound covalently. The subsequent primer extension reactions are performed, imaged, and washed with ammonia at each step until all five primers have been extended and the fluorescence incorporated by the primer has been imaged at each step.

The next step consists of determining a very short stretch of nucleotide sequence in the amplified DNA in each replica on the nitrocellulose. This entails sequencing just nine bases in a clone (which are referred to as the sequence tag of the clone). In this example, this is accomplished by two separate sets of sequencing reactions taking place in each replica-membrane. One set of sequencing reactions will determine the first five bases in the upper strand of the clone. The other set of sequencing reactions will determine the first five bases in the lower strand of the clone (always reading 5' to 3') as shown by the underlined X's below.

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 5' - NNNNNNNNNNNNNNNXXXXXXXXNNNNNNNNNNNNNNNNNNNN - 3'
 3' - NNNNNNNNNNNNNNNXXXXXXXXNNNNNNNNNNNNNNNNNNNN - 5'
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Each stretch of five bases is interrogated by using a mixture of specific primers for each base to be sequenced, using a single addition of a dideoxynucleotide triphosphate (ddNTP). The 3' end of the first primer (Primer 1) is positioned just before the first base to be sequenced. The sequence of the primer is defined by complementarity to vector sequences flanking the insert (or non-variable sequences flanking the region to be sequenced). The 3' end of the second primer (Primer 2) is positioned just before the second base to be sequenced. The sequence of the Primer 2 is defined by complementarity to the flanking sequences, but the last base at the 3' end is degenerate. An example design for Primer 1, Primer 2, and subsequent primers is shown below. The letter N indicates interrogation bases in a clone. The letter D indicates a degenerate base position in the primer. The question mark (?) indicates the nucleotide to be added to the primer.

Cloned sequence (SEQ ID NO:1):

TAAGTCTAGTTGACAGGATGCATGNNNNNNNNNtcagacagttgttgactgatggctg
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

Primer 1 (complexity = 1) (SEQ ID NOs:2 and 1)

TCTAGTTGACAGGATGCATG?
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

Primer 2 (complexity = 4) (SEQ ID NOs:3 and 1)

CTAGTTGACAGGATGCATGD?
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

Primer 3 (complexity = 16) (SEQ ID NOs:4 and 1)

TAGTTGACAGGATGCATGDD?
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

Primer 4 (complexity = 64) (SEQ ID NOs:5 and 1)

AGTTGACAGGATGCATGDDD?
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

Primer 5 (complexity = 256) (SEQ ID NOs:6 and 1)

GTTGACAGGATGCATGDDDD?
ATTCAGATCAACTGTCCTACGTACNNNNNNNNNagtcctgtcaacaactgactaccgac

While primer 1 is not degenerate, primer 2 contains one degenerate position, primer 3 contains two degenerate positions, primer 4 contains three degenerate positions, and primer 5 contains four degenerate positions.

Although primers 4 and 5 may prime at incorrect positions, the low complexity of the amplified DNA in a DNA colony produced by the disclosed method tends to ensure correct priming reads, on the average.

Primer extension is carried out for 5 minutes at 38°C in a primer extension solution containing Primer 1 as the only primer. The primer extension mixture preferably contains a thermostable DNA polymerase such as Taq polymerase, and a mixture of four fluorescent dideoxy-oligodeoxyribonucleotides, each labeled with a different dye, as in standard fluorescent sequencing (Perkin Elmer-Applied Biosystems, Inc.). Because only dideoxynucleotides are present in each colony replica, and because the colony contains millions of copies of the nucleic acid sequence of interest, the added fluorescent label will be easily detectable for each reaction. After

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 5'-NNNNNNNNNNNNNNNNXXXXXXXXXX...//.XXXXXXXXXXNNNNNNNNNNNNNNNNNN-3'
 3'-NNNNNNNNNNNNNNNNXXXXXXXXXX...//.XXXXXXXXXXNNNNNNNNNNNNNNNNNN-5'
 <<<<

5 The short five-base sequences on each end, together with the sequences of
flanking restriction enzyme sites, are sufficient to serve the function of
unique tagging of each cDNA clone using the procedures described above.

10 Replica-binding of amplified nucleic acid molecules to glass slides,
5 as described elsewhere herein, enables sequencing using a single slide. This
is accomplished by washing the slide with 1% ammonia after imaging of the
first primer extension reaction. This alkaline solution denatures the labeled
15 primer, but the cloned nucleic acid remains on the slide because it is bound
covalently. The subsequent primer extension reactions are performed,
20 imaged, and washed with ammonia at each step until all five primers have
been extended and the fluorescence incorporated by the primer has been
imaged at each step.

An alternative method to read the output of nanosequencing reactions
is to use mass spectroscopy instead of fluorescence. The use of mass
15 spectroscopy for sequence identification of primers that have been extended
by only one base has been described by Haff and Smirnov, *Genome*
25 *Research* 7: 378-388 (1997).

The sequencing scheme shown above can permit the sequencing of a
30 population of cDNA molecules derived from a single type of mRNA
20 molecules. This can be accomplished, for example, as follows. First, a
specific mRNA is amplified from any biological source using RT-PCR to
obtain full-length amplification products. The amplified PCR product may
35 have been derived from a mixture of wild-type sequence transcripts and also
a small proportion (1/100, for example) of mutant transcripts that contain a
25 single point mutation at a specific locus. Any other DNA fragment can also
be used.

40 The PCR product is then nicked with DNase I to generate a random
population of DNA fragments. The DNA is nick-translated with Klenow
DNA polymerase to generate a population of DNA fragments in the range of
45 120 to 200 nucleotides. This population of subsequences generated from the
30 population of cDNA is then cloned into linear vectors to form circular
vectors and the strands of the circular vectors separated as described herein.
50 Vectors with inserts in the size range of 120 to 200 base pairs can be isolated

(preferably before strand separation) by gel electrophoresis, or, preferably, by chromatography in Mono-Q 5/5/ (Pharmacia-LKB). The circular vectors are then amplified as described herein and the sequence of the ends of the inserts (which are thus the sequence tags) is determined as described above.

The sequence tags obtained from each clone consist of two pentamer (or even two hexamer) sequences. These sequence tags, which are known to be separated by segments of 120 to 200 bases, are catalogued and assembled into a contiguous sequence using techniques developed for hybridization sequencing. Starting with a cDNA product of a single type of mRNA, the cDNA can be entirely sequenced by assembling a catalog of sequenced clones. The sequence obtained from each clone is a pair of non-adjacent pentamers or hexamers. When a large number of molecular clones are analyzed, the method can reveal the presence of point mutations, even if they are present as 1/100th of the cDNA population.

Starting with a complex mixture of cDNAs, small DNA segments (sequence tags) present in clones that originated from individual cDNA molecules in the cDNA population can be sequenced *in situ* using a similar procedure. The method can be scaled up by increasing the density of molecular colonies, and the number of colony replicas.

I. Illustrations of the Method

The disclosed method is further illustrated by the following examples.

Illustration 1: Cloning using a linker

A DNA sample is amplified by PCR using standard procedures, except that both oligonucleotide primers are designed to contain unique restriction enzyme sites, such that after amplification the PCR product may be cleaved, generating different sticky ends on each side of the linear DNA product. One of the PCR primers additionally contains a spacer sequence. The digested PCR product is then placed in a ligation mixture containing linkers designed to circularize the amplified DNA. The linkers represent the linear vector. The linkers are designed with a chemically modified terminus in one of the oligonucleotides, such that after ligation the resulting circular DNA molecules (that is circular vectors) will contain a single nick (or

several nicks, if more than one linker is incorporated by ligation) in one of the strands. The modifier group may be a biotin, and it may be located either at the 5' or the 3' end of one of the linker oligonucleotides.

One of the PCR primers can also contain additional non-priming sequence designed to constitute the small spacer or backbone of the circular vector to allow amplification by rolling circle replication. The spacer sequence preferably contains a site for a rare-cutter restriction enzyme, which can be used to regenerate circles from linear DNA produced by amplification.

Optionally, the continuous strand of the circular vector can be separated from the discontinuous strand and unligated vector pieces using a two-step procedure. First, the circular vector, which contains a biotin residue, is bound to beads containing streptavidin, in order to bind the vector via the biotin present in one of the DNA strands that comprise the circular vector. The beads are then washed with formamide at mildly alkaline pH. Under appropriate conditions, the circularized DNA, which contains an unligated nick site by design, separates into two DNA molecules. Thus, mild alkaline-formamide denaturation releases free single-stranded circles from the beads. The single-stranded circular molecules are then further purified by gel filtration or ion exchange (Mono-Q 5/5) chromatography in the presence of an alkaline buffer (15 mM NaOH). This purification step will remove small linear molecules that contaminate the circular vector (which contain inserted nucleic acid molecules). The purpose of the purification procedure is useful for selecting certain DNA size classes, because this is desirable in certain applications. This separation is optional and the procedure can be performed with out strand separation or purification.

Dilutions of the DNA are then mixed with a buffer containing two primers (at approximately 1 μ Molar concentration) designed for strand displacement cascade amplification (that is, a secondary strand displacement primer and a rolling circle replication primer/tertiary strand displacement primer) and melted agarose at 60°C. The solution is overlayed on a petri dish to form a thin agarose layer (1.0% to 2% agarose), such that the concentration of circular DNA molecules is in the range of 500 to 5,000 per

5 plate. Then enzymes and dNTPs required to initiate rolling circle replication
are then added. The agarose film is incubated for 0.5 to 3 hours at 38°C (if
the enzyme used is exo-Klenow) or at 60°C (if the enzyme used is exo (-)
Bst or exo (-) Bca; Walker *et al.* (1992); Walker, IBC International
10 5 Conference, December, 1996). At appropriate dilutions, molecular
"colonies" clonally derived from single circular vectors bearing DNA that
originated in the PCR-amplified product, will form in the thin film of
agarose. The initial density of seed DNA molecules should be such that the
15 molecular colonies do not overlap.

10 In a preferred embodiment, one of the primers used in the
amplification reaction is capable of forming a secondary structure. By
lowering the temperature from 60°C to 50°C, this special primer forms a
hairpin structure that interferes with priming while the other primer
continues to function normally. As the amplification reaction is continued
15 for another 45 minutes, a large proportion of the DNA product becomes
single-stranded DNA generated by strand displacement driven by the single
functional primer. In this way, during the latter phase of the amplification
reaction a large proportion of the DNA contained in each colony becomes
single-stranded.
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20 After amplification, the agarose film is placed in a vacuum-blotting
device with a membrane that contacts the bottom of the agarose film. Part of
the DNA on the agarose film is vacuum-blotted onto the membrane. This
generates a replica of the DNA colonies on the surface of the membrane.
35 Blotting is carried out for a brief period of time, so that approximately half of
the amplified DNA remains in the agarose.
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40 In a preferred embodiment, the DNA from the colonies may be
blotted to a CAM membrane, a special membrane that permits reversible
binding of DNA. CAM is cellulose acetate membrane containing cystamine
(2,2'-dithio-bis[ethylamine]). The membrane contains primary amino
45 30 groups, positively charged below pH 9.5, that can be easily removed under
mild reductive conditions. CAM has been used to reversibly capture DNA
fragments separated by electrophoresis. CAM has been successfully used
with DNA fragments ranging from 0.5 to 320 Kbp. CAMs with different
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group densities can be synthesized (up to 1.65 $\mu\text{mole/sq cm}$); CAM with 1 $\mu\text{mole amino/sq cm}$ has a binding capacity of at least 10 $\mu\text{g DNA/sq cm}$. The standard elution conditions for DNA fragments up to 10 Kbp are: 2 hours at room temperature in 25 mM EDTA, 0.2 M NaCl, and 25 mM 2-mercaptoethanol. Larger fragments require higher concentrations of the reducing agent. The chemistry involved in the preparation of CAM is well established (see Sundberg and Porath, *J. Chromatog.* 90:87-98 (1974); Uy and Wolf, (1977)).

CAM is prepared in two steps: (1) Oxirane groups are introduced by reacting cellulose acetate membranes (0.45 μm) with variable concentrations (0 to 30 % v:v; depending upon the final group density required) of 1,4-butanediol diglycidyl ether in 0.1 M NaOH containing 2 mg/ml sodium borohydride; the reaction is allowed to proceed for 16 hours at RT, with mild agitation. (2) Cystamine is then coupled to the oxirane-containing membrane by reacting with 0.1 M cystamine in 0.1 M sodium tetraborate buffer, pH 9.5, for 16 hours at 37°C. Newly synthesized CAM is fully stable for at least 120 days at 4°C. The content of both oxirane and amino groups can be easily determined by standard reactions.

The thin agarose gel containing amplified DNA molecular colonies may be stained with a sensitive dye such as SIBR-GREEN II (Molecular Probes) in order to localize the position of each colony. The coordinates of the colony position then serves to locate the position of the replicas on the membrane.

In order to recover DNA from a CAM membrane and obtain single-stranded DNA that can be sequenced by standard methods, the procedure is as follows: Molecular colonies are generated as described above to generate single-stranded DNA (embodiment using one specifically structured primer that is inactivated by lowering the temperature), blotted to a CAM membrane, then a small droplet of DNA elution buffer (25 mM EDTA, 0.2 M NaCl, and 25 mM 2-mercaptoethanol) is placed on top of the desired colony, releasing in a few minutes a large proportion of the DNA of that colony replica. The small droplet is then recovered and mixed with four

volumes of a buffer containing a sequencing primer and a suitable sequencing mixture for standard Sanger dideoxy sequencing.

In order to recover DNA from a membrane and regenerate replicatable DNA circles that can be amplified in solution by rolling circle replication, or grown again as molecular colonies, the procedure is as follows: a small droplet of elution buffer (25 mM EDTA, 0.2 M NaCl, and 25 mM 2-mercaptoethanol) is placed on top of the desired colony, releasing in a few minutes a large proportion of the DNA of that colony. The small droplet is then recovered and mixed with four volumes of a buffer containing a restriction enzyme that will cleave the amplified DNA at the rare-cutter site that was designed into the spacer sequence of one of the original PCR primers. After inactivating the restriction enzyme, the DNA is treated very briefly with highly diluted alkaline phosphatase, in order to cause partial dephosphorylation of the termini of the cleaved DNA. After phosphatase inactivation, the DNA is diluted and ligated in the presence of T4 DNA ligase, thus regenerating closed circular molecules. A fraction of the recircularized molecules will contain a single nick, resulting from dephosphorylated ligation junctions, and these molecules will be capable of initiating rolling circle replication and strand displacement cascade amplification reactions.

Illustration 2: Cloning using a Y-vector

1. Vector design: This linear vector has a 3' protruding T residue at each end, so as to permit ligation with PCR products that contain a 3'-terminal A (generated during PCR) at each end. The panhandle or tail of the Y is formed by two oligonucleotides that together constitute the second strand of the linear vector. The longer oligonucleotide contains an oligo-dA sequence of 16 bases at the 3' terminus. The oligo-dA sequence serves as an affinity tag (where the affinity target will be oligo-dT). The shorter oligonucleotide contains a 5' phosphate. Sequences of an example of a functional Y-vector are shown below.

K.58 (SEQ ID NO:7)

P-CATGAGGACTAGCAGATGGATGCGGCCGAGCTCG

TGTAATACGACTCACTATAGGGT-3'

A.60 (SEQ ID NO:8)

P-CCCTATAGTGAGTCGTATTACACGAGCTGCTAGCAT
CATTAGCCAAAAAAAAAAAAAAAAA-3

B.42 (SEQ ID NO:9)

5 P-GGCTAATGATGCTAGGCCGCATCCATCTGCTAGTCCTCATGT-3'

2. The Y-vector is assembled by incubation of oligonucleotide K.58, A.60 and B.42, for 5 minutes at 40°C, and then ligated with a mixture of PCR-amplicons using T4 DNA ligase at 16°C for 16 hours, to generate circular vectors with inserts. Oligonucleotide K.58 is in the first strand (that is, continuous strand) of the circular vector. Oligonucleotides A.60 and B.42 are in the second strand (that is, the discontinuous strand) of the circular vector. Insert sequences are in both strands of the circular vector.

3. After ligation, the vectors with ligated inserts are incubated at room temperature with oligo-dT-cellulose (Life Sciences, Inc.) in the presence of DNA ligase. The oligo-dT-cellulose is an affinity substrate where the cellulose is the solid-state substrate and the oligo-dT is the affinity target. The Y-vector is ligated to the solid matrix via the panhandle sequence (the 5' end of B.42 is covalently bound to the 3' end of the oligo-dT on the oligo-dT-cellulose). The solid matrix is then washed with 20 mM Tris pH 8, 0.1 M NaCl, to remove unligated vectors.

4. The matrix is washed with 0.5 ml of 50 mM NaOH, releasing single-stranded circular DNA (the first strand of the circular vector) from the cellulose matrix. The now immobilized second strand of the circular vector remains attached to the cellulose matrix.

5. The circular vector is diluted serially to obtain concentrations in the range of several million circular vector molecules per milliliter. Approximately 22 µl of this DNA solution (containing approximately 30,000 to 120,000 DNA molecules) is placed on a cover slip, and covered with a polylysine-coated microscope slide. The DNA is allowed to bind to the polylysine-covered surface for 30 minutes at 37°C. The slide then dipped in 0.01% Tween-20, and dried at room temperature.

5 6. Using an arraying instrument, an array is constructed consisting of
6000 individual micro-droplets of a solution containing two suitable primers
designed for the constant sequence domains of the Y-vector, compatible
buffer, and polymerase (Large fragment Bst, or exo-Vent DNA polymerase)
10 5 capable of supporting rolling circle replication.

Primer 1 (23) (SEQ ID NO:10)

GCATCCATCTGCTAGTCCTCATG

Primer 2 (22) (SEQ ID NO:11)

CGCAGCTCGTGTAAATACGACTC

15 10 Primer 1 serves as the rolling circle replication primer and a tertiary
strand displacement primer. Primer 2 serves as a secondary strand
displacement primer. The use of these primers will result in strand
20 displacement cascade amplification. The diameter of the droplets should be
approximately 0.150 to 0.200 millimeters. Droplets are dispensed on the
15 surface of the slide in a controlled humidity atmosphere, in order to maintain
a constant droplet volume for a period of 90 minutes. Alternatively, the
25 diluted circular DNA molecules may be placed on the glass surface using the
arrayer instrument to dispense small volumes of liquid that on the average
contain a single ligated DNA molecule.

30 20 7. The array is incubated for 90 minutes at constant temperature (62°C)
to amplify any DNA molecules in contact with (or within) the droplets.
When 33% of the droplets grow DNA colonies, and the rest do not grow
35 anything, most colonies are likely to be of clonal origin. Adjusting the initial
inoculum density, it should be possible to obtain up to 1500 clonal colonies
25 per 6000-droplet array.

40 Optionally, the amplified nucleic acid can be replica plated in order
to save a copy of the clones or to perform additional operations on the
clones. In order to facilitate replica plating (via covalent binding to a glass
surface), each of the two primers used for the strand displacement cascade
45 30 amplification of the circular vectors may be synthesized with a primary
amino group at the 5' end. At the end of the SDCA reaction, a glass slide is
placed over the glass slide with the reaction droplets leaving an air gap of
less than 1 mm (defined by the thickness of a plastic spacer) in such a
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5 manner that the glass on top will contact all of the liquid droplets without
excessive compression. Prior to use, the lower face of this slide (the upper
slide) is derivatized using the methods described by Guo *et al.*, *Nucleic Acids*
10 5 *Research* 22;5456-5465 (1994), and Guo *et al.*, *Nature Biotechnology*
15:331-335 (1997)), to make the glass surface chemically reactive with
amino groups. The two-slide sandwich is incubated for 1 to 2 hours at 37°C
in order to obtain covalent coupling of the amplified DNA contained in each
droplet to the lower face of the upper slide.

8. DNA colonies may be identified by staining with the dye Sybr-
10 Green-I (Molecular Probes, Inc.). Alternatively, replica slides may be made
as indicated above, and used for any desired microarray hybridization
experiment. The DNA in each colony may also be isolated and identified or
20 analyzed by DNA sequencing.

Claims

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I claim:

1. A method of isolating and amplifying a nucleic acid molecule, the method comprising
 - (a) ligating a nucleic acid molecule into a linear vector to form a circular vector comprising the vector and the nucleic acid molecule, wherein the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, wherein the second strand of the circular vector is discontinuous, and wherein the first strand in the circular vector is a closed circular strand,
 - (b) amplifying the first strand by rolling circle replication to form tandem sequence DNA, wherein the amplification results in amplification of the nucleic acid molecule in the first strand.
2. The method of claim 1 wherein the second strand of the linear vector contains at least one nick, wherein the nick cannot be ligated.
3. The method of claim 1 wherein either the 5' or the 3' end of the second strand of the linear vector cannot be ligated.
4. The method of claim 1 wherein the second strand of the linear vector contains at least one gap or overlap.
5. The method of claim 1 wherein the method further comprises, following ligation and prior to amplification, separating the first strand from the second strand.
6. The method of claim 5 wherein the second strand includes an affinity tag.
7. The method of claim 6 wherein the first strand is separated from the second strand by binding the affinity tag to a substrate, denaturing the first and second strands prior to, simultaneous with, or following binding, and separating the first strand from the substrate.
8. The method of claim 5 wherein the second strand of the linear vector contains at least one overlap, part of the overlapping portions of the second strand are complementary, and the 3' end of the overlap extends beyond the part of the overlapping portions that are complementary,

5 wherein the first strand is separated from the second strand by ligating
one end of the second strand to a nucleic acid molecule coupled to a substrate,
denaturing the first and second strands following ligation of the second strand,
and separating the first strand from the substrate.

10 9. The method of claim 1 wherein step (a) comprises ligating a plurality
of nucleic acid molecules into a plurality of linear vectors in a single reaction to
form a plurality of circular vectors, each circular vector containing at least one
nick, gap, or overlap in the second strand,

15 wherein step (b) comprises amplifying the first strand of the plurality of
circular vectors, and

20 wherein the method further comprises, prior to amplification, dividing the
ligation reaction to produce a plurality of separate amplification reactions.

10. The method of claim 9 further comprising making a replica of the
amplification reactions.

25 11. The method of claim 10 wherein the replica of the amplification
reactions is made by contacting the amplification reactions with a surface to
which nucleic acids can bind.

30 12. The method of claim 10 wherein the replica of the amplification
reactions is made by transferring part of each amplification reaction to form a
replica amplification reaction.

35 13. The method of claim 9 wherein the ligation reaction is divided by
spreading the ligation reaction onto a surface to form a spread, and wherein the
separate amplification reactions are the locations of circular vectors on the
surface after spreading.

40 14. The method of claim 13 further comprising making a replica of the
amplification reactions.

15. The method of claim 14 wherein the replica is made by contacting the
spread with a second surface to which nucleic acids can bind.

45 16. The method of claim 9 wherein any number or all of the amplification
reactions are ordered as an array of reaction droplets or in an array of reaction
vessels.

5 17. The method of claim 16 wherein, following amplification, all or part of the contents of any number or all the individual reaction droplets or reaction vessels are transferred by one to one mapping to a new set of reaction droplets or reaction vessels.

10 18. The method of claim 17 further comprising, following amplification, determining the presence of amplified nucleic acid in the amplification reactions, and

15 transferring all or a part of the contents of the amplification reactions containing amplified nucleic acid reaction to a new set of reaction droplets or reaction vessels.

20 19. The method of claim 16 further comprising making a replica of the amplification reactions.

 20. The method of claim 19 wherein the replica of the amplification reactions is made by contacting the amplification reactions with a surface to which nucleic acids can bind.

25 21. The method of claim 19 wherein the replica of the amplification reactions is made by contacting the amplification reactions with a surface treated with an affinity target capable of binding an affinity tag, wherein the amplified nucleic comprises affinity tags incorporated during amplification,

30 wherein a portion of each amplification reaction is transferred to the surface.

35 22. The method of claim 21 wherein the affinity tag is biotin and the affinity target is streptavidin.

40 23. The method of claim 21 wherein the affinity tag is a reactive moiety and the affinity target is a corresponding reactive moiety, where a chemical reaction between the affinity tag and the affinity target results in the amplified nucleic acid being covalently coupled to the surface.

45 24. The method of claim 23 wherein the affinity target is phenylene diisothiocyanate, disuccinimidylcarbonate, disuccinimidyl oxalate or dimethylsuberimidate and the affinity tag is a reactive amine.

50 25. The method of claim 19 wherein the replica of the amplification reactions is made by transferring part of each amplification reaction to form a replica amplification reaction.

5 26. The method of claim 16 wherein, following amplification, all or part
of the contents of any number or all of the reaction droplets or reaction vessels
are transferred and combined to create one or more sets of pooled reactions.

10 27. The method of claim 16 wherein the amplification reactions are
arranged on the surface of a substrate.

15 28. The method of claim 27 wherein the substrate comprises acrylamide,
cellulose, nitrocellulose, polystyrene, polyethylene vinyl acetate, polypropylene,
polymethacrylate, polyethylene, polyethylene oxide, glass, polysilicates,
polycarbonates, teflon, fluorocarbons, nylon, silicon rubber, polyanhydrides,
polyglycolic acid, polylactic acid, polyorthoesters, polypropylfumerate, collagen,
glycosaminoglycans, polyamino acids, chemical resistant metals, or corrosion
20 resistant metals.

29. The method of claim 9 wherein the method further comprises, prior to
dividing the ligation reaction,

25 diluting the ligation reaction such that, on average, each amplification
reaction contains a single circular vector.

30 30. The method of claim 9 wherein the method further comprises,
following amplification, collecting a sample of each amplification reaction.

31. The method of claim 9 wherein the method further comprises
35 detecting or sequencing the nucleic acid molecules in the amplification reactions
or in the collected samples.

32. The method of claim 1 wherein rolling circle replication is primed by
35 the second strand.

33. The method of claim 1 wherein rolling circle replication is primed by
a rolling circle replication primer.

40 34. The method of claim 33 wherein the tandem sequence DNA is
amplified by strand displacement replication to form secondary tandem sequence
DNA.

45 35. The method of claim 34 wherein the secondary tandem sequence
DNA is amplified by strand displacement replication to form tertiary tandem
sequence DNA.

50 36. The method of claim 34 wherein strand displacement replication of
the tandem sequence is primed by a strand displacement primer.

5 37. The method of claim 9 further comprising detecting one or more amplified nucleic acid molecules in one or more of the amplification reactions.

38. The method of claim 37 wherein the nucleic acid molecules are derived from cDNA generated by suppression subtractive hybridization.

10 39. The method of claim 37 wherein the plurality of nucleic acid molecules are all derived from the same source.

40. The method of claim 37 further comprising, following amplification, creating a replica of the amplification reactions, contacting the amplification reactions with a first set of labeled nucleic acid probes and the replica amplification reactions with a second set of labeled nucleic acid probes, and

20 comparing the pattern of hybridization of the first set of probes to the pattern of hybridization of the second set of probes, wherein differences in the patterns of hybridization indicate differences in the probe sets.

25 41. The method of claim 40 further comprising selecting for isolation or further analysis amplification reactions that hybridize to the first set of probes but not to the second set of probes, amplification reactions that hybridize to the second set of probes but not to the first set of probes, amplification reactions that hybridize to the both sets of probes, or amplification reactions that do not hybridize to either set of probes.

30 42. An *in vitro* method of cloning nucleic acid molecules, the method comprising

35 (a) dividing a nucleic acid sample to produce a plurality of separate amplification reactions,

40 (b) amplifying nucleic acid molecules in the amplification reactions,

(c) making a replica of the amplification reactions,

(d) testing nucleic acid molecules in either the amplification reactions or the replica amplification reactions to identify nucleic acid molecules of interest, and

45 (e) retrieving the identified nucleic acid molecules of interest from the corresponding amplification reactions or replica amplification reactions that were not tested.

5 43. The method of claim 42 wherein the nucleic acid sample is divided
by spreading the sample onto a surface to form a spread, and wherein the separate
amplification reactions are the locations of circular vectors on the surface after
spreading.

10 44. The method of claim 43 wherein the replica of the amplification
reactions is made by contacting the spread with a second surface to which nucleic
acids can bind.

15 45. The method of claim 42 wherein the method further comprises, prior
to dividing the nucleic acid sample,

diluting the nucleic acid sample such that, on average, each amplification
reaction contains a single nucleic acid molecule.

20 46. A method of isolating and amplifying nucleic acid molecules, the
method comprising

(a) ligating a plurality of nucleic acid molecules into a plurality of linear
vectors in a single reaction to form a plurality of circular vectors, each circular
25 vector comprising a vector and a nucleic acid molecule,

wherein the linear vectors are double-stranded linear nucleic acid
comprising two nucleic acid strands, wherein the circular vectors each contain at
least one nick, gap, or overlap in the second strand, and wherein the first strand in
30 each circular vector is a closed circular strand,

(b) separating the first strands from the second strands,

(c) diluting and dividing the first strands to produce a plurality of
35 separate amplification reactions that, on average, each contain a single circular
vector,

(d) amplifying the first strands of the plurality of circular vectors by
rolling circle replication to form tandem sequence DNA,

40 wherein the amplification results in amplification of the nucleic acid
molecules in the first strands.

45 47. The method of claim 46 wherein the tandem sequence DNA is
amplified by strand displacement replication to form secondary tandem sequence
DNA.

5 48. The method of claim 47 wherein the secondary tandem sequence DNA is amplified by strand displacement replication to form tertiary tandem sequence DNA.

10 49. A method of isolating and amplifying a nucleic acid molecule, the method comprising

 (a) ligating a nucleic acid molecule into a linear vector to form a circular vector comprising the vector and the nucleic acid molecule,

15 wherein the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, wherein the circular vector contains at least one nick in the second strand and wherein the first strand in the circular vector is a closed circular strand,

20 (b) amplifying the first strand,

 wherein the amplification results in amplification of the nucleic acid molecule in the first strand.

25 50. A kit for isolating and amplifying nucleic acid molecules, the kit comprising

 (a) a linear vector wherein the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, and wherein

30 (1) the linear vector contains at least one nick, wherein the nick cannot be ligated,

 (2) either the 5' or the 3' end of the second strand of the linear vector cannot be ligated,

35 (3) the second strand of the linear vector contains at least one gap,

 (4) the second strand of the linear vector contains at least one overlap, or

40 (5) any combination of (1), (2), (3) or (4);

 (b) a rolling circle replication primer, wherein the rolling circle replication primer is complementary to a portion of the first strand of the linear vector; and

45 (c) a strand displacement primer, wherein the strand displacement primer matches a portion of the first strand of the linear vector.

5 51. A linear vector wherein the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, wherein the second strand of the linear vector contains an affinity tag, and wherein

10 (1) the linear vector contains at least one nick, wherein the nick cannot be ligated,

(2) either the 5' or the 3' end of the second strand of the linear vector cannot be ligated,

15 (3) the second strand of the linear vector contains at least one gap,

(4) the second strand of the linear vector contains at least one overlap, or

(5) any combination of (1), (2), (3) or (4).

20 52. A linear vector wherein the linear vector is a double-stranded linear nucleic acid comprising two nucleic acid strands, and wherein the second strand of the linear vector contains at least one overlap, part of the overlapping portions of the second strand are complementary, and the 3' end of the overlap extends
25 beyond the part of the overlapping portions that are complementary.

Linear vectors

Circular vectors

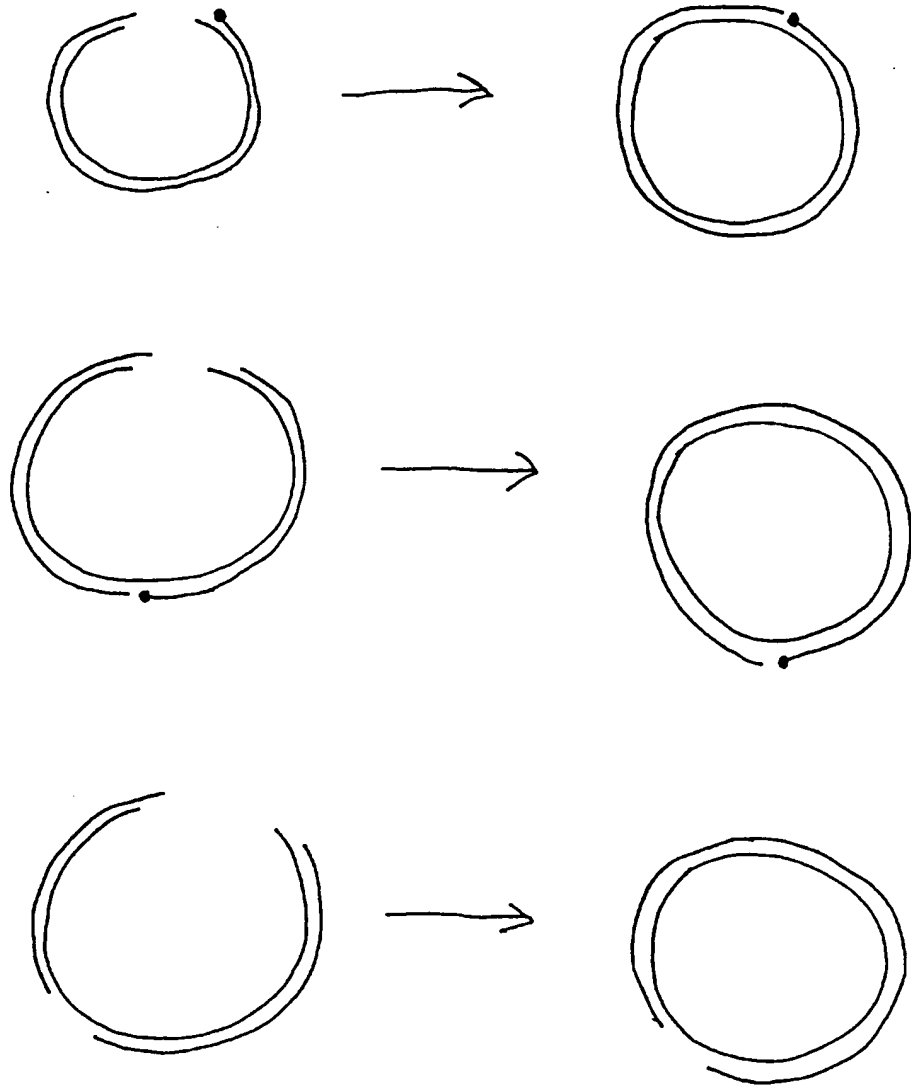


Figure 1A

Figure 1B

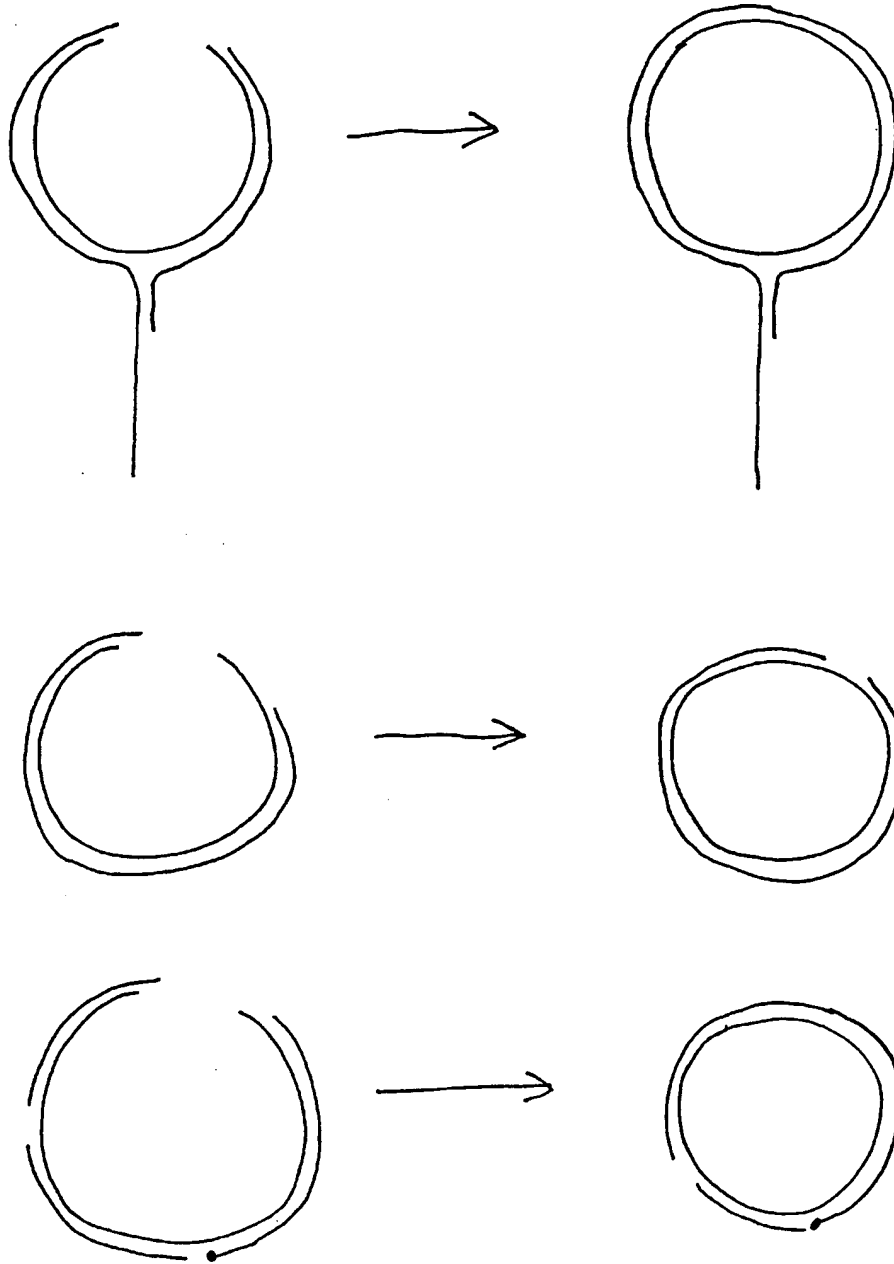


Figure 1C

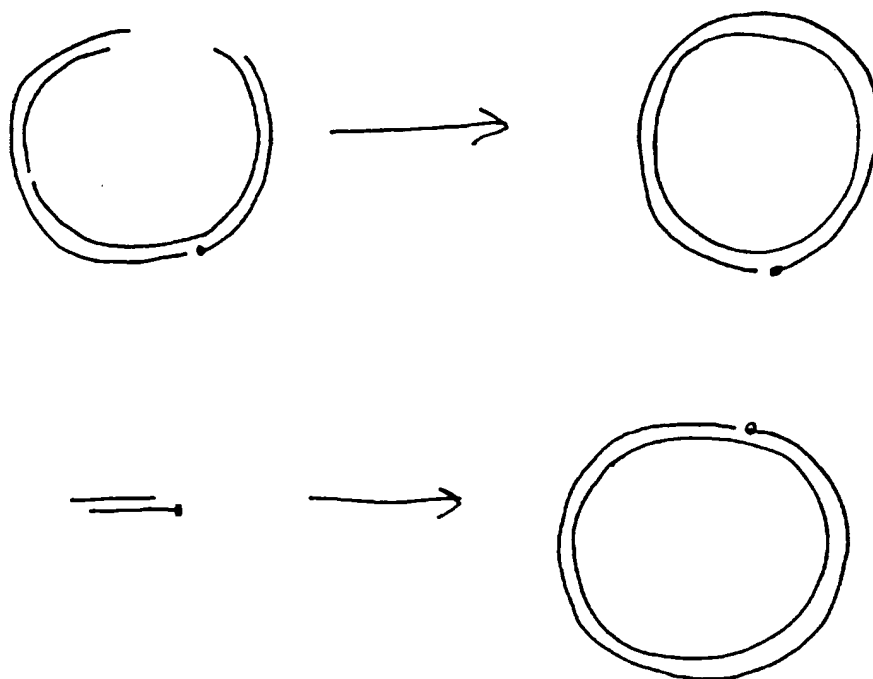


Figure 2A

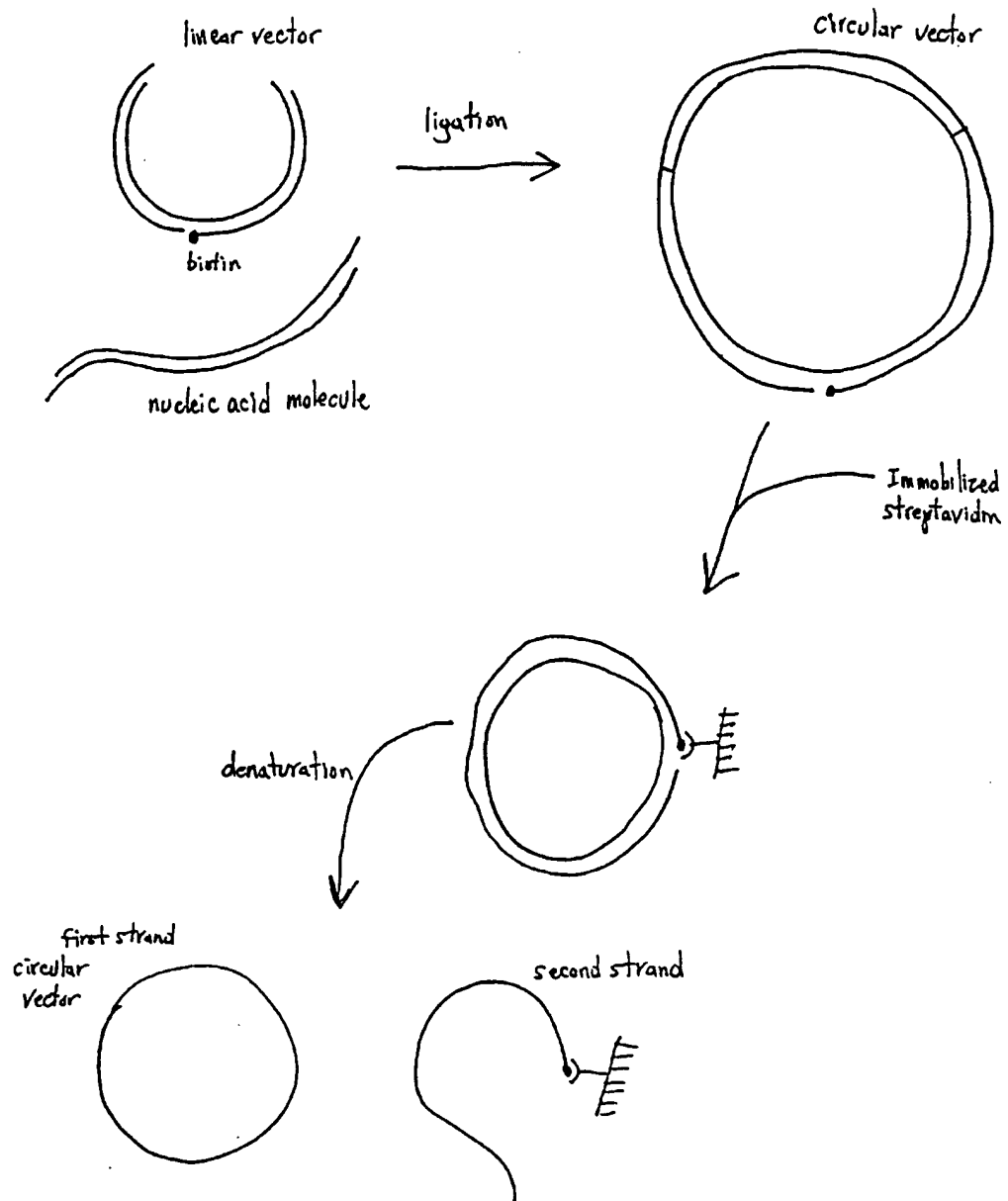
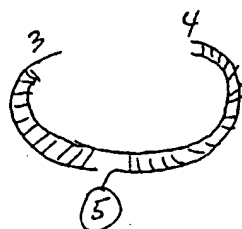


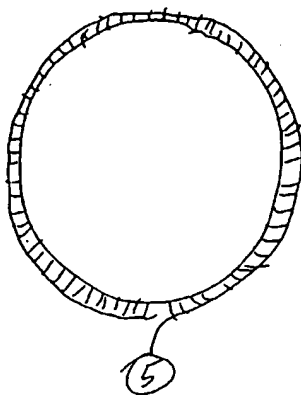
Figure 2B



Vector

1, 3 are compatible
sticky ends

2, 4 are blunt
ends



Insert
has been
ligated
into vector

5 is biotin
or any
hapten

Figure 3

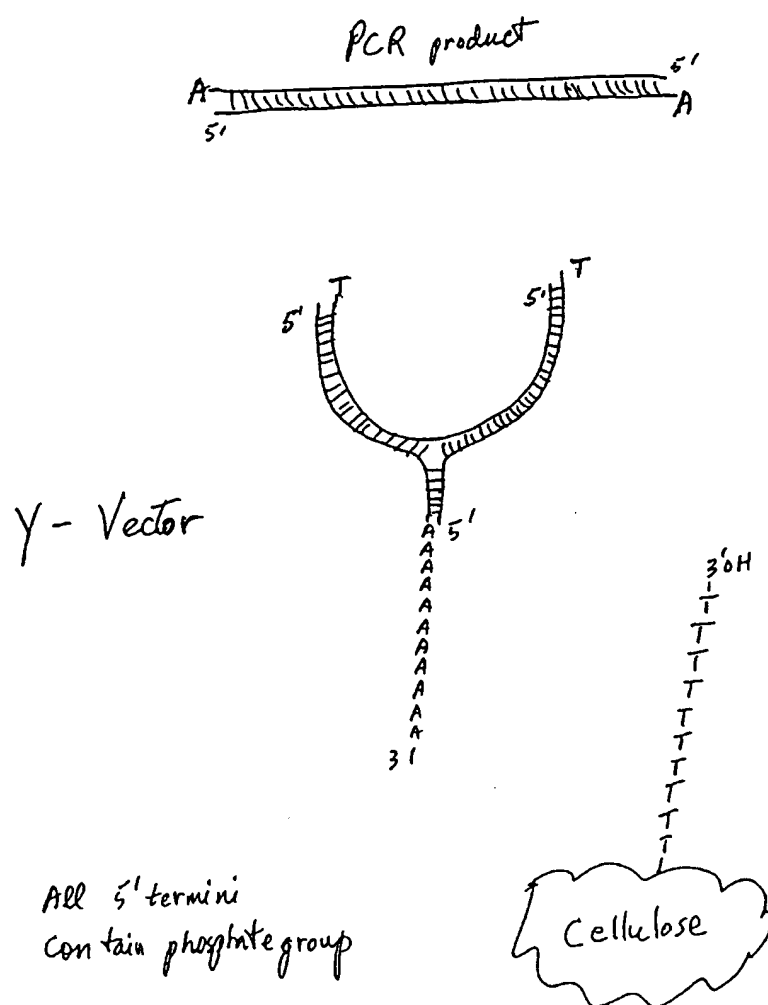
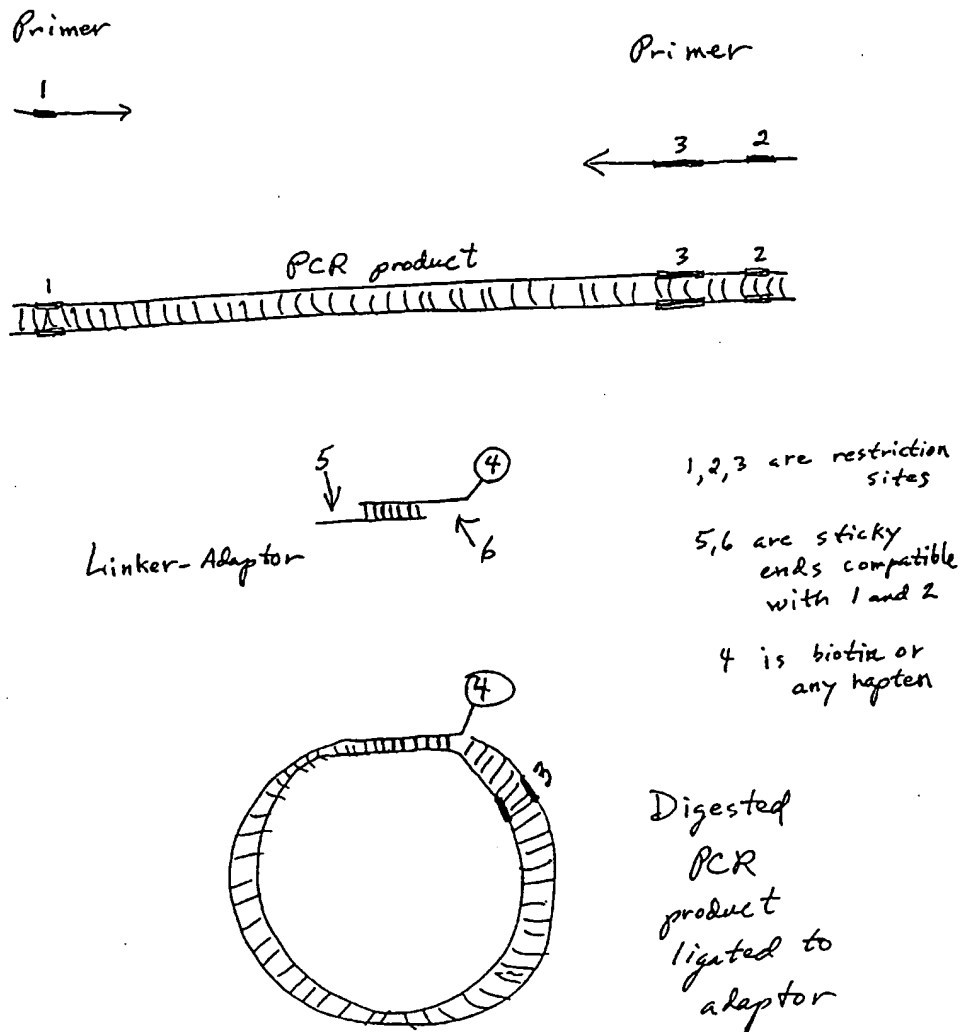


Figure 4





INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification: C12Q 1/68	A3	(11) International Publication Number: WO 00/15779
		(43) International Publication Date: 23 March 2000 (23.03.2000)

(21) International Application Number: PCT/US99/21291

(22) International Filing Date: 15 September 1999 (15.09.1999)

(30) Priority Data:
60/100,327 15 September 1998 (15.09.1998) US(60) Parent Application or Grant
YALE UNIVERSITY [/]; (). LIZARDI, Paul, M. [/];
(). PABST, Patrea, L.; ().

Published

(54) Title: MOLECULAR CLONING USING ROLLING CIRCLE AMPLIFICATION

(54) Titre: CLONAGE MOLECULAIRE A AMPLIFICATION SELON LE MODELE DU CERCLE ROULANT

(57) Abstract

Disclosed are reagents and a method for efficient in vitro molecular cloning of nucleic acid molecules of interest. Because the method is entirely in vitro, it can be automated and scaled-up in ways that are not possible in cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process. The amplification is rapid and efficient since it involves a single, isothermic reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be used for any purpose and in any manner that nucleic acid cloned or amplified by known methods can be used. This includes sequencing, probing, restriction analysis, subcloning, transcription, hybridization or denaturation analysis, further amplified, and storage for future use or analysis.

(57) Abrégé

L'invention concerne des réactifs et un procédé permettant d'assurer un clonage moléculaire in vitro de molécules d'acides nucléiques déterminées. Etant entièrement in vitro, le procédé peut être automatisé et étendu selon des modalités qu'il est impossible de mettre en oeuvre avec le clonage moléculaire à base cellulaire. Le procédé consiste à insérer la molécule d'acide nucléique concernée dans un vecteur linéaire, de manière à former un vecteur circulaire dans lequel un brin est continu et l'autre brin est discontinu. Le brin continu est ensuite amplifié selon le modèle du cercle roulant, ce qui permet d'amplifier par la même occasion la molécule d'acide nucléique. L'amplification est rapide et efficace car elle fait intervenir une réaction isothermique unique qui assure la réplication exponentielle des séquences du vecteur. Le procédé peut être automatisé lorsque plusieurs réactions sont conduites simultanément dans une zone de taille réduite. L'acide nucléique amplifié peut être utilisé à tout fin et de façon quelconque dans l'éventail d'utilisation connu de l'acide nucléique cloné ou amplifié, y compris le séquençage, la détermination de sondes, l'analyse de restriction, le sous-clonage, la transcription, l'analyse d'hybridation ou de dénaturation, le complément d'amplification et le stockage pour utilisation ou analyse ultérieure.

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12Q 1/68	A3	(11) International Publication Number: WO 00/15779 (43) International Publication Date: 23 March 2000 (23.03.00)
(21) International Application Number: PCT/US99/21291 (22) International Filing Date: 15 September 1999 (15.09.99) (30) Priority Data: 60/100,327 15 September 1998 (15.09.98) US (71) Applicant: YALE UNIVERSITY [US/US]; 451 College Street, New Haven, CT 06511 (US). (72) Inventor: LIZARDI, Paul, M.; 350 Mountain Road, Hamden, CT 06514 (US). (74) Agents: PABST, Patrea, L. et al.; Amall Golden & Gregory, LLP, 2800 One Atlantic Center, 1201 West Peachtree Street, Atlanta, GA 30309-3450 (US).	(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published With international search report. (88) Date of publication of the international search report: 10 August 2000 (10.08.00)	
(54) Title: MOLECULAR CLONING USING ROLLING CIRCLE AMPLIFICATION (57) Abstract Disclosed are reagents and a method for efficient <i>in vitro</i> molecular cloning of nucleic acid molecules of interest. Because the method is entirely <i>in vitro</i> , it can be automated and scaled-up in ways that are not possible in cell-based molecular cloning. The method involves insertion of a nucleic acid molecule of interest in a linear vector to form a circular vector where one strand is continuous and the other strand is discontinuous. The continuous strand of the circular vector is then amplified by rolling circle replication, amplifying the inserted nucleic acid molecule in the process. The amplification is rapid and efficient since it involves a single, isothermic reaction that replicates the vector sequences exponentially. The amplification process is amenable to automation where multiple reactions are carried out simultaneously in a small area. The amplified nucleic acid can be used for any purpose and in any manner that nucleic acid cloned or amplified by known methods can be used. This includes sequencing, probing, restriction analysis, subcloning, transcription, hybridization or denaturation analysis, further amplified, and storage for future use or analysis.		

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INTERNATIONAL SEARCH REPORT

 International Application No.
 PCT/US 99/21291

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C12Q1/68		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 7 C12Q		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
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